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(54) Title: BMP-9 COMPOSITIONS

TGA + -41	Thr					TGT Cys						48
						ACA Thr						96
						Arg					CAC His	144
						AAC Asn						192
						TAT Tyr					GGC Gly	240
						gat Asp					GCT Ala 55	. 288
											aag Lys	336
				Pro		AGC Ser 80	Pro				AAG Lys	384
			Gly			Lys				Gly	AGC Ser	432
		Glu				TATC	TGC	CTGC	GGG			470

(57) Abstract

Purified BMP-9 proteins and processes for producing them are disclosed. The proteins may be used in the treatment of bone and cartilage defects and in wound healing and related tissue repair.

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BMP-9 COMPOSITIONS

The present invention relates to a novel family of purified proteins designated BMP-9 proteins and processes for obtaining them. These proteins may be used to induce bone and/or cartilage formation and in wound healing and tissue repair.

The murine BMP-9 DNA sequence (SEQ ID NO: 1) and amino acid sequence (SEQ ID NO: 2) are set forth in Figure 1. Human BMP-9 sequence is set forth in Figure 3 (SEQ ID NO: 8 and SEQ ID NO: 9). It is contemplated that BMP-9 proteins are capable of inducing the formation of cartilage and/or bone. BMP-9 proteins may be further characterized by the ability to demonstrate cartilage and/or bone formation activity in the rat bone formation assay described below.

Murine BMP-9 is characterized by comprising amino acid #319 to #428 of Figure 1 (SEQ ID NO: 2 amino acids #1-110). Murine BMP-9 may be produced by culturing a cell transformed with a DNA sequence comprising nucleotide #610 to nucleotide #1893 as shown in Figure 1 (SEQ ID NO: 1) and recovering and purifying from the culture medium a protein characterized by the amino acid sequence comprising amino acid #319 to #428 as shown in Figure-1 (SEQ ID NO: 2) substantially free from other proteinaceous materials with which it is co-produced.

Human BMP-9 is expected to be homologous to murine BMP-9 and is characterized by comprising amino acid #1 (Ser, Ala, Gly) to #110 of Figure 3 (SEQ ID NO: 9) (Arg). The invention includes methods for obtaining the DNA sequences encoding human BMP-9. This method entails utilizing the murine BMP-9 nucleotide sequence or portions thereof to design probes to screen libraries for the human gene or fragments thereof using standard techniques. Human BMP-9 may be produced by culturing

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a cell transformed with the BMP-9 DNA sequence and recovering and purifying BMP-9 from the culture medium. The expressed protein is isolated, recovered, and purified from the culture medium. The purified expressed protein is substantially free from other proteinaceous materials with which it is coproduced, as well as from other contaminants. The recovered purified protein is contemplated to exhibit cartilage and/or bone formation activity. The proteins of the invention may be further characterized by the ability to demonstrate cartilage and/or bone formation activity in the rat bone formation assay described below.

Human BMP-9 may be produced by culturing a cell transformed with a DNA sequence comprising nucleotide #124 to #453 as shown in SEQ ID NO: 8 and recovering and purifying from the culture medium a protein characterized by the amino acid sequence of SEQ ID NO: 9 from amino acid #1 to amino acid #110 substantially free from other proteinaceous materials with which it is co-produced.

Another aspect of the invention provides pharmaceutical compositions containing a therapeutically effective amount of a BMP-9 protein in a pharmaceutically acceptable vehicle or carrier. BMP-9 compositions of the invention may be used in the formation of cartilage. These compositions may further be utilized for the formation of bone. BMP-9 compositions may also be used for wound healing and tissue repair. Compositions of the invention may further include at least one other therapeutically useful agent such as the BMP proteins BMP-1, BMP-2, BMP-3, BMP-4, BMP-5, BMP-6, and BMP-7 disclosed for instance in PCT publications WO88/00205, WO89/10409, and WO90/11366, and BMP-8, disclosed in U.S. application Ser. No. 07/641,204 filed January 15, 1991, Ser. No. 07/525,357 filed May 16, 1990, and Ser. No. 07/800,364 filed November 20, 1991.

The compositions of the invention may comprise, in addition to a BMP-9 protein, other therapeutically useful agents including growth factors such as epidermal growth factor (EGF), fibroblast growth factor (FGF), transforming growth

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factor (TGF- α and TGF- β), and insulin-like growth factor (IGF). The compositions may also include an appropriate matrix for instance, for supporting the composition and providing a surface for bone and/or cartilage growth. The matrix may provide slow release of the osteoinductive protein and/or the appropriate environment for presentation thereof.

The BMP-9 compositions may be employed in methods for treating a number of bone and/or cartilage defects, periodontal disease and various types of wounds. These methods, according to the invention, entail administering to a patient needing such bone and/or cartilage formation wound healing or tissue repair, an effective amount of a BMP-9 protein. These methods may also entail the administration of a protein of the invention in conjunction with at least one of the novel BMP proteins disclosed in the co-owned applications described above. In addition, these methods may also include the administration of a BMP-9 protein with other growth factors including EGF, FGF, $TGF-\alpha$, $TGF-\beta$, and IGF.

Still a further aspect of the invention are DNA sequences coding for expression of a BMP-9, protein. Such sequences include the sequence of nucleotides in a 5' to 3' direction illustrated in Figure 1 (SEQ ID NO: 1) and Figure 3 (SEQ ID NO: 8) or DNA sequences which hybridize under stringent conditions with the DNA sequences of Figure 1 or 3 and encode a protein having the ability to induce the formation of cartilage and/or bone. Finally, allelic or other variations of the sequences of Figure 1 or 3, whether such nucleotide changes result in changes in the peptide sequence or not, are also included in the present invention.

A further aspect of the invention includes vectors comprising a DNA sequence as described above in operative association with an expression control sequence therefor. These vectors may be employed in a novel process for producing a BMP-9 protein of the invention in which a cell line transformed with a DNA sequence encoding a BMP-9 protein in

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operative association with an expression control sequence therefor, is cultured in a suitable culture medium and a BMP-9 protein is recovered and purified therefrom. This process may employ a number of known cells both prokaryotic and eukaryotic as host cells for expression of the polypeptide.

Other aspects and advantages of the present invention will be apparent upon consideration of the following detailed description and preferred embodiments thereof.

10 Brief Description of the Drawing

FIG. 1 comprises DNA sequence and derived amino acid sequence of murine BMP-9 from clone ML14a further described below.

FIG. 2 comprises DNA sequence and derived amino acid sequence of human BMP-4 from lambda U2OS-3 ATCC #40342.

FIG. 3 comprises DNA sequence and derived amino acid sequence of human BMP-9 from λ FIX/H6111 ATCC # 75252.

20 <u>Detailed Descripton of the Invention</u>

The murine BMP-9 nucleotide sequence (SEQ ID NO: 1) and encoded amino acid sequence (SEQ ID NO: 2) are depicted in Figure 1. Purified murine BMP-9 proteins of the present invention are produced by culturing a host cell transformed wth a DNA sequence comprising the DNA coding sequence of Figure 1 (SEQ ID NO: 1) from nucleotide #610 to nucleotide #1893 and recovering and purifying from the culture medium a protein which contains the amino acid sequence or a substantially homologous sequence as represented by amino acid #319 to #428 of Figure 1 (SEQ ID NO: 2). The BMP-9 proteins recovered from the culture medium are purified by isolating them from other proteinaceous materials from which they are co-produced and from other contaminants present.

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Human BMP-9 nucleotide and amino acid sequence is depicted in SEQ ID No: 8 and 9. Mature human BMP-9 is expected to comprise amino acid #1 (Ser, Ala, Gly) to #110 (Arg).

Human BMP-9 may be produced by culturing a cell transformed with a DNA sequence comprising nucleotide #124 to #453 as shown in SEQ ID NO: 8 and recovering and purifying from the culture medium a protein characterized by the amino acid sequence of SEQ ID NO: 9 from amino acid #1 to amino acid #110 substantially free from other proteinaceous materials with which it is co-produced.

BMP-9 proteins may be characterized by the ability to induce the formation of cartilage. BMP-9 proteins may be further characterized by the ability to induce the formation of bone. BMP-9 proteins may be further characterized by the ability to demonstrate cartilage and/or bone formation activity in the rat bone formation assay described below.

The BMP-9 proteins provided herein also include factors encoded by the sequences similar to those of Figure 1 and 3 (SEQ ID NO's: 1 and 8), but into which modifications are naturally provided (e.g. allelic variations in the nucleotide sequence which may result in amino acid changes polypeptide) or deliberately engineered. For example, synthetic polypeptides may wholly or partially duplicate continuous sequences of the amino acid residues of Figure 1 of Figure 3 (SEQ ID No's: 2 and 9). These sequences, by virtue of sharing primary, secondary, or tertiary structural conformational characteristics with bone growth polypeptides of Figure 1 and Figure 3 may possess bone growth factor biological properties in common therewith. Thus, they employed as biologically active substitutes naturally-occurring BMP-9 and other BMP-9 polypeptides in therapeutic processes.

Other specific mutations of the sequences of BMP-9 proteins described herein involve modifications of glycosylation sites. These modifications may involve O-linked

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or N-linked glycosylation sites. For instance, the absence of glycosylation or only partial glycosylation results from amino deletion at asparagine-linked substitution or glycosylation recognition sites. The asparagine-linked glycosylation recognition sites comprise tripeptide sequences which are specifically recognized by appropriate cellular glycosylation enzymes. These tripeptide sequences are either asparagine-X-threonine or asparagine-X-serine, where X is usually any amino acid. A variety of amino acid substitutions or deletions at one or both of the first or third amino acid positions of a glycosylation recognition site (and/or amino acid deletion at the second position) results in nonglycosylation at the modified tripeptide sequence.

The present invention also encompasses the novel DNA sequences, free of association with DNA sequences encoding other proteinaceous materials, and coding on expression for BMP-9 proteins. These DNA sequences include those depicted in Figure 1 or Figure 3 (SEQ ID NO's: 1 and 8) in a 5' to 3' direction and those sequences which hybridize thereto under stringent hybridization conditions [see, T. Maniatis et al, Molecular Cloning (A Laboratory Manual), Cold Spring Harbor Laboratory (1982), pages 387 to 389] and encode a protein having cartilage and/or bone inducing activity.

Similarly, DNA sequences which code for BMP-9 proteins coded for by the sequences of Figure 1 or Figure 3, but which differ in codon sequence due to the degeneracies of the genetic code or allelic variations (naturally-occurring base changes in the species population which may or may not result in an amino acid change) also encode the novel factors described herein. Variations in the DNA sequences of Figure 1 or Figure 3 (SEQ ID NO: 1 and 8) which are caused by point mutations or by induced modifications (including insertion, deletion, and substitution) to enhance the activity, half-life or production of the polypeptides encoded are also encompassed in the invention.

Another aspect of the present invention provides a novel

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method for producing BMP-9 proteins. The method of the present invention involves culturing a suitable cell line, which has been transformed with a DNA sequence encoding a BMP-9 protein of the invention, under the control of known regulatory sequences. The transformed host cells are cultured and the BMP-9 proteins recovered and purified from the culture medium. The purified proteins are substantially free from other proteins with which they are co-produced as well as from other contaminants.

Suitable cells or cell lines may be mammalian cells, such as Chinese hamster ovary cells (CHO). The selection of suitable mammalian host cells and methods for transformation, culture, amplification, screening, product production and purification are known in the art. See, e.g., Gething and Sambrook, Nature, 293:620-625 (1981), or alternatively, Kaufman et al, Mol. Cell. Biol., 5(7):1750-1759 (1985) or Howley et al, U.S. Patent 4,419,446. Another suitable mammalian cell line, which is described in the accompanying examples, is the monkey COS-1 cell line. The mammalian cell CV-1 may also be suitable.

Bacterial cells may also be suitable hosts. For example, the various strains of \underline{E} . $\underline{\operatorname{coli}}$ (e.g., HB101, MC1061) are well-known as host cells in the field of biotechnology. Various strains of \underline{B} . $\underline{\operatorname{subtilis}}$, $\underline{\operatorname{Pseudomonas}}$, other bacilli and the like may also be employed in this method.

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Many strains of yeast cells known to those skilled in the art may also be available as host cells for expression of the polypeptides of the present invention. Additionally, where desired, insect cells may be utilized as host cells in the method of the present invention. See, e.g. Miller et al, Genetic Engineering, 8:277-298 (Plenum Press 1986) and references cited therein.

Another aspect of the present invention provides vectors for use in the method of expression of these novel BMP-9 polypeptides. Preferably the vectors contain the full novel

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DNA sequences described above which encode the novel factors of Additionally the vectors also contain invention. appropriate expression control sequences permitting expression of the BMP-9 protein sequences. Alternatively, vectors incorporating modified sequences as described above are also embodiments of the present invention. The vectors may be employed in the method of transforming cell lines and contain selected regulatory sequences in operative association with the DNA coding sequences of the invention which are capable of directing the replication and expression thereof in selected host cells. Regulatory sequences for such vectors are known to those skilled in the art and may be selected depending upon the host cells. Such selection is routine and does not form part of the present invention.

protein of the present invention, which induces cartilage and/or bone formation in circumstances where bone is not normally formed, has application in the healing of bone fractures and cartilage defects in humans and other animals. Such a preparation employing a BMP-9 protein may have prophylactic use in closed as well, as open fracture reduction and also in the improved fixation of artificial joints. novo bone formation induced by an osteogenic agent contributes to the repair of congenital, trauma induced, or oncologic resection induced craniofacial defects, and also is useful in cosmetic plastic surgery. A BMP-9 protein may be used in the treatment of periodontal disease, and in other tooth repair. processes. Such agents may provide an environment to attract bone-forming cells, stimulate growth of bone-forming cells or induce differentiation of progenitors of bone-forming cells. BMP-9 polypeptides of the invention may also be useful in the osteogenic, treatment of osteoporosis. A variety of cartilage-inducing and bone inducing factors have been described. See, e.g. European patent applications 148;155 and 169,016 for discussions thereof.

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The proteins of the invention may also be used in wound healing and related tissue repair. The types of wounds include, but are not limited to burns, incisions and ulcers. (See, e.g. PCT Publication WO84/01106 for discussion of wound healing and related tissue repair).

It is further contemplated that proteins of the invention may increase neuronal survival and therefore be useful in transplantation and treatment of conditions exhibiting a decrease in neuronal survival.

A further aspect of the invention is a therapeutic method and composition for repairing fractures and other conditions related to cartilage and/or bone defects or periodontal diseases. The invention further comprises therapeutic methods and compositions for wound healing and tissue repair. Such compositions comprise a therapeutically effective amount of at least one of the BMP-9 proteins of the invention in admixture with a pharmaceutically acceptable vehicle, carrier or matrix.

It is expected that the proteins of the invention may act in concert with or perhaps synergistically with other related proteins and growth factors. Further therapeutic methods and compositions of the invention therefore comprise a therapeutic amount of at least one BMP-9 protein of the invention with a therapeutic amount of at least one of the other BMP proteins disclosed in co-owned applications described above. combinations may comprise separate molecules of the BMP proteins or heteromolecules comprised of different For example, a method and composition of the invention may comprise a disulfide linked dimer comprising a BMP-9 protein subunit and a subunit from one of the "BMP" proteins described above. A further embodiment may comprise a heterodimer of BMP-9 moieties. Further, BMP-9 proteins may be combined with other agents beneficial to the treatment of the bone and/or cartilage defect, wound, or tissue in question. These agents include various growth factors such as epidermal growth factor (EGF), platelet derived growth factor (PDGF),

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transforming growth factors (TGF- α and TGF- β), and insulin-like growth factor (IGF).

The preparation and formulation of such physiologically acceptable protein compositions, having due regard to pH, isotonicity, stability and the like, is within the skill of the art. The therapeutic compositions are also presently valuable for veterinary applications due to the lack of species specificity in BMP proteins. Particularly domestic animals and thoroughbred horses in addition to humans are desired patients for such treatment with BMP-9 of the present invention.

The therapeutic method includes administering composition topically, systemically, or locally as an implant or device. When administered, the therapeutic composition for use in this invention is, of course, in a pyrogen-free, physiologically acceptable form. Further, the composition may desirably be encapsulated or injected in a viscous form for delivery to the site of bone, cartilage or tissue damage. -Topical administration may be suitable for wound healing and tissue repair. Therapeutically useful agents other than the BMP-9 proteins which may also optionally be included in the above, may alternatively composition as described additionally, be administered simultaneously or sequentially with the BMP composition in the methods of the invention.

Preferably for bone and/or cartilage formation, the composition would include a matrix capable of delivering BMP-9 or other BMP proteins to the site of bone and/or cartilage damage, providing a structure for the developing bone and cartilage and optimally capable of being resorbed into the body. The matrix may provide slow release of BMP-9 and/or the appropriate environment for presentation thereof. Such matrices may be formed of materials presently in use for other implanted medical applications.

The choice of matrix material is based on biocompatibility, biodegradability, mechanical properties, cosmetic appearance and interface properties. The particular

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compositions will define application of the BMP-9 the appropriate formulation. Potential matrices for compositions may be biodegradable and chemically defined calcium sulfate, tricalciumphosphate, hydroxyapatite, polylactic acid and polyanhydrides. Other potential materials are biodegradable and biologically well defined, such as bone or dermal collagen. Further matrices are comprised of pure proteins or extracellular matrix components. Other potential matrices are nonbiodegradable and chemically defined, such as sintered hydroxyapatite, bioglass, aluminates, ceramics. Matrices may be comprised of combinations of any of the above mentioned types of material, such as polylactic acid and hydroxyapatite or collagen and tricalciumphosphate. bioceramics may be altered in composition, such as in calciumaluminate-phosphate and processing to alter pore size, particle size, particle shape, and biodegradability.

The dosage regimen will be determined by the attending physician considering various factors which modify the action of the BMP-9 protein, e.g. amount of bone weight desired to be formed, the site of bone damage, the condition of the damaged bone, the size of a wound, type of damaged tissue, the patient's age, sex, and diet, the severity of any infection, time of administration and other clinical factors. The dosage may vary with the type of-matrix used in the reconstitution and the types of BMP proteins in the composition. The addition of other known growth factors, such as IGF I (insulin like growth factor I), to the final composition, may also effect the Progress can be monitored by periodic assessment of bone growth and/or repair, for example, x-rays, histomorphometric determinations and tetracycline labeling.

The following examples illustrate practice of the present invention in recovering and characterizing murine BMP-9 protein and employing it to recover the human and other BMP-9 proteins, obtaining the human proteins and expressing the proteins via recombinant techniques.

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EXAMPLE I

Murine BMP-9

750,000 recombinants of a mouse liver cDNA library made in the vector lambdaZAP (Stratagene/Catalog #935302) are plated and duplicate nitrocellulose replicas made. A fragment of human BMP-4 DNA corresponding to nucleotides 1330-1627 of Figure 2 (SEQ ID NO: 3) (the human BMP-4 sequence) is ^{12}P labeled by the random priming procedure of Feinberg et al. [Anal. Biochem. 132: 6-13 (1983)] and hybridized to both sets of filters in SHB at 60°C for 2 to 3 days. Both sets of filters are washed under reduced stringency conditions (4X SSC, 0.1% SDS at 60°C). Many duplicate hybridizing recombinants of various intensities (approximately 92) are noted. 50 of the strongest hybridizing recombinant bacteriophage are plaque purified and their inserts are transferred to the plasmid Bluescript SK (+/-) according to the in vivo excision protocol described by the manufacturer (Stratagene). DNA sequence -analysis-of-several recombinants indicate that they encode a protein homologous to other BMP proteins and other proteins in the TGF- β family. The DNA sequence and derived amino acid sequence of one recombinant, designated ML14a, is set forth in Figure 1. (SEQ ID NO: 1)

The nucleotide sequence of clone ML14a contains an open reading frame of 1284 bp, encoding a BMP-9 protein of 428 amino acids. The encoded 428 amino acid BMP-9 protein is contemplated to be the primary translation product as the coding sequence is preceded by 609 bp of 5' untranslated sequence with stop codons in all three reading frames. The 428 amino acid sequence predicts a BMP-9 protein with a molecular weight of 48,000 daltons.

Based on knowledge of other BMP proteins and other proteins within the TGF- β family, it is predicted that the precursor polypeptide would be cleaved at the multibasic sequence ARG-ARG-LYS-ARG in agreement with a proposed consensus

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proteolytic processing sequence of ARG-X-X-ARG. Cleavage of the BMP-9 precursor polypeptide at this location would generate a 110 amino acid mature peptide beginning with the amino acid SER at position #319. The processing of BMP-9 into the mature form is expected to involve dimerization and removal of the N-terminal region in a manner analogous to the processing of the related protein $TGF-\beta$ [L.E. Gentry, et al., Molec. & Cell. Biol. 8:4162 (1988); R. Derynck, et al., Nature 316:701 (1985)].

It is contemplated therefore that the mature active species of murine BMP-9 comprises a homodimer of 2 polypeptide subunits, each subunit comprising amino acids #319-#428 with a predicted molecular weight of approximately 12,000 daltons. Further active species are contemplated comprising amino acids #326 - #428 thereby including the first conserved cysteine residue. As with other members of the BMP and $TGF-\beta$ family of proteins, the carboxy-terminal region of the BMP-9 protein exhibits greater sequence conservation than the more aminoterminal portion. The percent amino acid identity of the murine BMP-9 protein in the cysteine-rich C-terminal domain (amino acids #326 - #428) to the corresponding region of other human BMP proteins and other proteins within the TGF- β family is as follows: BMP-2, 53%; BMP-3, 43%; BMP-4, 53%; BMP-5, 55%; BMP-6, 55%; BMP-7, 53%; Vgl, 50%; GDF-1, 43%; TGF- β 1, 32%; TGF- β 2, 34%; TGF- β 3, 34%; inhibin β (B), 34%; and inhibin β (A), 42%.

EXAMPLE II

Human BMP-9

Murine and human osteoinductive factor genes are presumed to be significantly homologous, therefore the murine coding sequence or a portion thereof is used as a probe to screen a human genomic library or as a probe to identify a human cell line or tissue which synthesizes the analogous human cartilage and/or bone protein. A human genomic library (Toole et al., supra) may be screened with such a probe, and presumptive

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positives isolated and DNA sequence obtained. Evidence that this recombinant encodes a portion of the human BMP-9 relies of the murine/human protein and gene structure homologies.

Once a recombinant bacteriophage containing DNA encoding portion of the human cartilage and/or bone inductive factor molecule is obtained, the human coding sequence can be used as a probe to identify a human cell line or tissue which synthesizes BMP-9. Alternatively, the murine coding sequence can be used as a probe to identify such human cell line or tissue. Briefly described, RNA is extracted from a selected cell or tissue source and either electrophoresed on a formaldehyde agarose gel and transferred to nitrocellulose, or reacted with formaldehyde and spotted on nitrocellulose directly. The nitrocellulose is then hybridized to a probe derived from a coding sequence of the murine or human BMP-9. mRNA is selected by oligo (dT) cellulose chromatography and cDNA is synthesized and cloned in lambda gt10 or lambda ZAP by established techniques_(Toole_et al., supra).

Additional methods known to those skilled in the art may be used to isolate the human and other species' BMP-9 proteins of the invention.

A. Isolation of Human BMP-9 DNA

One million recombinants of a human genomic library constructed in the vector λFIX (Stratagene catalog # 944201) are plated and duplicate nitrocellulose replicas made. Two oligonucleotides probes designed on the basis of nucleotides #1665-#1704 and #1837-#1876 of the sequence set forth in Figure 1 (SEQ ID NO:1) are synthesized on an automated DNA synthesizer. The sequence of these two oligonucleotides is indicated below:

#1: CTATGAGTGTAAAGGGGGTTGCTTCTTCCCATTGGCTGAT

#2: GTGCCAACCCTCAAGTACCACTATGAGGGGATGAGTGTGG
These two oligonucleotide probes are radioactively labeled with

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 γ^{32} P-ATP and each is hybridized to one set of the duplicate nitrocellulose replicas in SHB at 65°C and washed with 1X SSC. 0.1% SDS at 65°C. Three recombinants which hybridize to both oligonucleotide probes are noted. All three positively hybridizing recombinants are plaque purified, bacteriophage plate stocks are prepared and bacteriophage DNA is isolated from each. The oligonucleotide hybridizing regions of one of these recombinants, designated HG111, is localized to a 1.2 kb Pst I/Xba I fragment. This fragment is subcloned into a plasmid vector (pGEM-3) and DNA sequence analysis is performed. HG111 was deposited with the ATCC, 12301 Parklawn Drive. Rockville, Maryland USA on June 16, 1992 under the requirements of the Budapest Treaty and designated as ATCC # 75252. subclone is designated pGEM-111. A portion of the DNA sequence of clone pGEM-111 is set forth in Figure 3 (SEQ ID NO:8/ HUMAN BMP-9 sequence). This sequence encodes the entire mature region of human BMP-9 and a portion of the propeptide. should be noted that this sequence consists of preliminary data. Particularly, the propeptide region is subject to further analysis and characterization. For example, nucleotides #1 through #3 (TGA) encode a translational stop which may be incorrect due to the preliminary nature of the sequence. It is predicted that additional sequences present in both pGEM-111 (the 1.2 kb PstI/XbaI fragment of HG111 subcloned into pGEM) and HG111 encode additional amino acids of the human BMP-9 propeptide region. Based on knowledge of other BMPs and other proteins within the TGF- β family, it is predicted that the precursor polypeptide would be cleaved at the multibasic sequence ARG-ARG-LYS-ARG (amino acids # -4 through # -1 of SEQUENCE ID NO:9) in agreement with a proposed consensus proteolytic processing sequence ARG-X-X-ARG. Cleavage of the human BMP-9 precursor polypeptide at this location would generate a 110 amino acid mature peptide beginning with the amino acid SER at position #1 of SEQUENCE ID NO:9 (encoded by

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nucleotides #124 through #126 of SEQUENCE ID NO:8). The processing of human BMP-9 into the mature form is expected to involve dimerization and removal of the N-terminal region in a manner analogous to the processing of the related protein $TGF-\beta$ [L.E. Gentry, et al., Molec. & Cell. Biol. 8:4162 (1988); R. Derynck, et al., Nature 316:701 (1985)].

It is contemplated therefore that the mature active species of human BMP-9 comprises a homodimer of two polypeptide subunits, each subunit comprising amino acids #1 through #110 of SEQUENCE ID NO:9, with a predicted molecular weight of Further active species are contemplated 12,000 daltons. comprising amino acids #8 through #110 thereby including the first conserved cysteine residue. As with other members of the BMP and TGF- β family of proteins, the carboxy-terminal portion the human BMP-9 sequence exhibits greater sequence of conservation than the amino-terminal portion. amino acid identity of the human BMP-9 protein in the cysteine-__rich_C-terminal_domain (amino acids #8 through #110) to the corresponding region of other human BMP proteins and other proteins within the TGF- β family is as follows: BMP-2, 52%; BMP-3, 40%; BMP-4, 52%; BMP-5, 55%; BMP-6, 55%; BMP-7, 53%; murine BMP-9, 97%; Vg1, 50%; GDF-1, 44%; TGF- β 1, 32%; TGF- β 2, 32%; TGF- β 3, 32%; inhibin β (B), 35%; and inhibin β (A), 41%.

25 EXAMPLE III

Rosen Modified Sampath-Reddi Assay

A modified version of the rat bone formation assay described in Sampath and Reddi, Proc. Natl. Acad. Sci. U.S.A., 80:6591-6595 (1983) is used to evaluate bone and/or cartilage activity of the BMP proteins. This modified assay is herein called the Rosen-modified Sampath-Reddi assay. The ethanol precipitation step of the Sampath-Reddi procedure is replaced by dialyzing (if the composition is a solution) or diafiltering (if the composition is a suspension) the fraction to be assayed against water. The solution or suspension is then redissolved

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in 0.1 % TFA, and the resulting solution added to 20mg of rat matrix. A mock rat matrix sample not treated with the protein serves as a control. This material is frozen and lyophilized and the resulting powder enclosed in #5 gelatin capsules. The capsules are implanted subcutaneously in the abdominal thoracic area of 21 - 49 day old male Long Evans rats. The implants are removed after 7 - 14 days. Half of each implant is used for alkaline phosphatase analysis [See, A. H. Reddi et al., Proc. Natl Acad Sci., 69:1601 (1972)].

The other half of each implant is fixed and processed for histological analysis. $1\mu m$ glycolmethacrylate sections are stained with Von Kossa and acid fuschin to score the amount of induced bone and cartilage formation present in each implant. The terms +1 through +5 represent the area of each histological section of an implant occupied by new bone and/or cartilage cells and matrix. A score of +5 indicates that greater than 50% of the implant is new bone and/or cartilage produced as a direct result of protein in the implant. A score of +4, +3, +2 and +1 would indicate that greater than 40%, 30%, 20% and 10% respectively of the implant contains new cartilage and/or bone. In a modified scoring method, three non-adjacent sections are evaluated from each implant and averaged. "+/-" indicates tentative identification cartilage or bone; "+1" indicates >10% of each section being new cartilage or bone; "+2", >25%; "+3", >50%; "+4", -75%; "+5", >80%. A "-" indicates that the implant is not recovered.

It is contemplated that the dose response nature of the BMP-9 containing samples of the matrix samples will demonstrate that the amount of bone and/or cartilage formed increases with the amount of BMP-9 in the sample. It is contemplated that the control samples will not result in any bone and/or cartilage formation.

As with other cartilage and/or bone inductive proteins such as the above-mentioned "BMP" proteins, the bone and/or cartilage formed is expected to be physically confined to the

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space occupied by the matrix. Samples are also analyzed by SDS gel electrophoresis and isoelectric focusing followed by autoradiography. The activity is correlated with the protein bands and pI. To estimate the purity of the protein in a particular fraction an extinction coefficient of 1 OD/mg-cm is used as an estimate for protein and the protein is run on SDS PAGE followed by silver staining or radioiodination and autoradiography.

10 EXAMPLE IV

Expression of BMP-9

In order to produce murine, human or other mammalian BMP-9 proteins, the DNA encoding it is transferred into an appropriate expression vector and introduced into mammalian cells or other preferred eukaryotic or prokaryotic hosts by conventional genetic engineering techniques. The preferred expression system for biologically active recombinant human BMP-9 is contemplated to be stably transformed mammalian cells.

One skilled in the art can construct mammalian expression vectors by employing the sequence of Figure 1 (SEQ ID NO: 1) or Figure 3 (SEQ ID NO: 8), or other DNA sequences encoding BMP-9 proteins or other modified sequences and known vectors, such as pCD [Okayama et al., Mol. Cell Biol., 2:161-170 (1982)], pJL3, pJL4 [Gough et al., EMBO J., 4:645-653 (1985)] and pMT2 CXM.

The mammalian expression vector pMT2 CXM is a derivative of p91023 (b) (Wong et al., Science 228:810-815, 1985) differing from the latter in that it contains the ampicillin resistance gene in place of the tetracycline resistance gene and further contains a XhoI site for insertion of cDNA clones. The functional elements of pMT2 CXM have been described (Kaufman, R.J., 1985, Proc. Natl. Acad. Sci. USA 82:689-693) and include the adenovirus VA genes, the SV40 origin of replication including the 72 bp enhancer, the adenovirus major late promoter including a 5' splice site and the majority of

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the adenovirus tripartite leader sequence present on adenovirus late mRNAs, a 3' splice acceptor site, a DHFR insert, the SV40 early polyadenylation site (SV40), and pBR322 sequences needed for propagation in $\underline{E.\ coli.}$

Plasmid pMT2 CXM is obtained by EcoRI digestion of pMT2-VWF, which has been deposited with the American Type Culture Collection (ATCC), Rockville, MD (USA) under accession number ATCC 67122. EcoRI digestion excises the cDNA insert present in pMT2-VWF, yielding pMT2 in linear form which can be ligated and used to transform E. coli HB 101 or DH-5 to ampicillin resistance. Plasmid pMT2 DNA can be prepared by conventional methods. pMT2 CXM is then constructed using loopout/in mutagenesis [Morinaga, et al., Biotechnology 84: 636 (1984). This removes bases 1075 to 1145 relative to the Hind III site near the SV40 origin of replication and enhancer sequences of pMT2. In addition it inserts the following sequence:

5' PO-CATGGGCAGCTCGAG-3' (SEQ ID NO: 5) at nucleotide 1145. This sequence contains the recognition site for the restriction endonuclease Xho I. A derivative of pMT2CXM, termed pMT23, contains recognition sites for the restriction endonucleases PstI, Eco RI, SalI and XhoI. Plasmid pMT2 CXM and pMT23 DNA may be prepared by conventional methods.

pEMC2bl derived from pMT21 may also be suitable in practice of the invention. pMT21 is derived from pMT2 which is derived from pMT2-VWF. As described above EcoRI digestion excises the cDNA insert present in pMT-VWF, yielding pMT2 in linear form which can be ligated and used to transform <u>E. Coli</u> HB 101 or DH-5 to ampicillin resistance. Plasmid pMT2 DNA can be prepared by conventional methods.

pMT21 is derived from pMT2 through the following two modifications. First, 76 bp of the 5' untranslated region of the DHFR cDNA including a stretch of 19 G residues from G/C tailing for cDNA cloning is deleted. In this process, a XhoI site is inserted to obtain the following sequence immediately

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upstream from DHFR: 5' -CTGCAGGCGAGCCTGAATTCCTCGAGCCATCATG-3'
PstI Eco RI XhoI
(SEQ ID NO: 6)

Second, a unique ClaI site is introduced by digestion with EcoRV and XbaI, treatment with Klenow fragment of DNA polymerase I, and ligation to a ClaI linker (CATCGATG). This deletes a 250 bp segment from the adenovirus associated RNA (VAI) region but does not interfere with VAI RNA gene expression or function. pMT21 is digested with EcoRI and XhoI, and used to derive the vector pEMC2B1.

A portion of the EMCV leader is obtained from pMT2-ECAT1 [S.K. Jung, et al, <u>J. Virol 63</u>:1651-1660 (1989)] by digestion with Eco RI and PstI, resulting in a 2752 bp fragment. This fragment is digested with TaqI yielding an Eco RI-TaqI fragment of 508 bp which is purified by electrophoresis on low melting agarose gel. A 68 bp adapter and its complementary strand are synthesized with a 5' TaqI protruding end and a 3' XhoI protruding end which has the following sequence:

20 5'-<u>CGA</u>GGTTAAAAAACGTCTAGGCCCCCGAACCACGGGGACGTGGTTTTCCTTT
TaqI

GAAAAACACG<u>ATT</u>GC-3' XhoI (SEQ ID NO: 7)

This sequence matches the EMC virus leader sequence from nucleotide 763 to 827. It also changes the ATG at position 10 within the EMC virus leader to an ATT and is followed by a XhoI site. A three way ligation of the pMT21 Eco RI-XhoI fragment, the EMC virus EcoRI-TaqI fragment, and the 68 bp

oligonucleotide adapter TaqI-XhoI adapter resulting in the vector pEMC2 β 1.

This vector contains the SV40 origin of replication and enhancer, the adenovirus major late promoter, a cDNA copy of the majority of the adenovirus tripartite leader sequence, a small hybrid intervening sequence, an SV40 polyadenylation

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signal and the adenovirus VA I gene, DHFR and β -lactamase markers and an EMC sequence, in appropriate relationships to direct the high level expression of the desired cDNA in mammalian cells.

The construction of vectors may involve modification of the BMP-9 DNA sequences. For instance, BMP-9 cDNA can be modified by removing the non-coding nucleotides on the 5' and 3' ends of the coding region. The deleted non-coding nucleotides may or may not be replaced by other sequences known to be beneficial for expression. These vectors are transformed into appropriate host cells for expression of BMP-9 proteins.

One skilled in the art can manipulate the sequences of Figure 1 or Figure 3 (SEQ ID NO: 1 and 8) by eliminating or replacing the mammalian regulatory sequences flanking the coding sequence with bacterial sequences to create bacterial vectors for intracellular or extracellular expression by bacterial cells. For example, the coding sequences could be further manipulated (e.g. ligated to other known linkers or modified by deleting non-coding sequences therefrom or altering nucleotides therein by other known techniques). The modified BMP-9 coding sequence could then be inserted into a known bacterial vector using procedures such as described in T. Taniguchi et al., Proc. Natl Acad. Sci. USA, 77:5230-5233 (1980).This exemplary bacterial vector could then be transformed into bacterial host cells and a BMP-9 protein expressed thereby. For a strategy for producing extracellular expression of BMP-9 proteins in bacterial cells, see, e.g. European patent application EPA 177,343.

Similar manipulations can be performed for the construction of an insect vector [See, e.g. procedures described in published European patent application 155,476] for expression in insect cells. A yeast vector could also be constructed employing yeast regulatory sequences intracellular or extracellular expression of the factors of the present invention by yeast cells. [See, e.g., procedures

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described in published PCT application WO86/00639 and European patent application EPA 123,289].

A method for producing high levels of a BMP-9 protein of the invention in mammalian cells may involve the construction of cells containing multiple copies of the heterologous BMP-9 gene. The heterologous gene is linked to an amplifiable marker, e.g. the dihydrofolate reductase (DHFR) gene for which cells containing increased gene copies can be selected for propagation in increasing concentrations of methotrexate (MTX) according to the procedures of Kaufman and Sharp, J. Mol. Biol., 159:601-629 (1982). This approach can be employed with a number of different cell types.

For example, a plasmid containing a DNA sequence for a BMP-9 of the invention in operative association with other plasmid sequences enabling expression thereof and the DHFR expression plasmid pAdA26SV(A)3 [Kaufman and Sharp, Mol. Cell. Biol., 2:1304 (1982)] can be co-introduced into DHFR-deficient CHO cells, DUKX-BII, by various methods including calcium phosphate coprecipitation and transfection, electroporation or protoplast fusion. DHFR expressing transformants are selected for growth in alpha media with dialyzed fetal calf serum, and subsequently selected for amplification by growth in increasing concentrations of MTX (e.g. sequential steps in 0.02, 0.2, 1.0 and 5uM MTX) as described in Kaufman et al., Mol Cell Biol., Transformants are cloned, and biologically 5:1750 (1983). active BMP-9 expression is monitored by the Rosen-modified Sampath - Reddi rat bone formation assay described above in Example III. BMP-9 expression should increase with increasing levels of MTX resistance. BMP-9 polypeptides are characterized using standard techniques known in the art such as pulse labeling with [35S] methionine or cysteine and polyacrylamide gel electrophoresis. Similar procedures can be followed to produce other related BMP-9 proteins.

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A. BMP-9 Vector Construction

In order to produce human BMP-9 proteins of the invention DNA sequences encoding the mature region of the human BMP-9 protein may be joined to DNA sequences encoding the propertide region of the murine BMP-9 protein. This murine/human hybrid DNA sequence is inserted into an appropriate expression vector and introduced into mammalian cells or other preferred eukaryotic or prokaryotic hosts by conventional genetic engineering techniques. The construction of this murine/human BMP-9 containing expression plasmid is described below.

A derivative of the human BMP-9 sequence (SEQ ID NO:8) comprising the nucleotide sequence from nucleotide #105 to #470 is specifically amplified. The following oligonucleotides are utilized as primers to allow the amplification of nucleotides #105 to #470 of the human BMP-9 sequence (SEQ ID NO:8) from clone pGEM-111 described above.

#3 ATCGGGCCCCTTTTAGCCAGGCGGAAAAGGAG

#4 AGCGAATTCCCCGCAGGCAGATACTACCTG

This procedure generates the insertion of the nucleotide sequence ATCGGGCCCCT immediately preceding nucleotide #105 and the insertion of the nucleotide sequence GAATTCGCT immediately following nucleotide #470. The addition of these sequences results in the creation of an Apa I and EcoR I restriction endonuclease site at the respective ends of the specifically amplified DNA fragment. The resulting 374 bp Apa I/EcoR I fragment is subcloned into the plasmid vector pGEM-72f(+) (Promega catalog# p2251) which has been digested with Apa I and EcoR I. The resulting clone is designated phBMP9mex-1.

The following oligonucleotides are designed on the basis of murine BMP-9 sequences (SEQ ID NO:1) and are modified to facilitate the construction of the murine/human expression plasmid referred to above:

#5

35 GATTCCGTCGACCACCATGTCCCCTGGGGCCTGGTCTAGATGGATACACAGCTGTGGGGCC

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#6 CCACAGCTGTGTATCCATCTAGACCAGGCCCCAGGGGACATGGTGGTCGACG
These oligonucleotides contain complimentary sequences which
upon addition to each other facilitate the annealing (base
pairing) of the two individual sequences, resulting in the
formation of a double stranded synthetic DNA linker (designated
LINK-1) in a manner indicated below:

This DNA linker (LINK-1) contains recognition sequences of restriction endonucleases needed to facilitate subsequent manipulations required to construct the murine/human expression plasmid, as well as sequences required for maximal expression of heterologous sequences in mammalian cell expression systems. More specifically (referring to the sequence numbering of #1-#11 oligonucleotide #5/LINK-1): nucleotides recognition sequences for the restriction endonucleases BamH I and Sal I, nucleotides #11-#15 allow for maximal expression of heterologuos sequences in mammallian cell expression systems, nucleotides #16-#31 correspond to nucleotides #610-#625 of the murine BMP-9 sequence (SEQ ID NO:1), nucleotides #32-#33 are inserted to facilitate efficient restriction digestion of two adjacent restriction endonuclease sites (EcoO109 I and Xba I), nucleotides #34-#60 correspond to nucleotides #1515-#1541 of the murine BMP-9 sequence (SEQ ID NO:1) except that nucleotide #58 of synthetic oligonucloetide #5 is a G rather than the A which appears at position #1539 of SEQ ID NO:1 (This nucleotide conversion results in the creation of an Apa I restriction

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endonuclease recognition sequence, without altering the amino acid sequence it is intended to encode, to facilitate further manipulations of the murine/human hybrid expression plasmid. LINK-1 (the double stranded product of the annealing of oligonucleotides #5 and #6) is subcloned into the plasmid vector pGEM-7Zf(+) which has been digested with the restriction endonucleases Apa I and BamH I. This results in a plasmid in which the sequences normally present between the Apa I and BamH I sites of the pGEM-7Zf(+) plasmid polylinker are replaced with the sequences of LINK-1 described above. The resulting plasmid clone is designated pBMP-9link.

pBMP-9link is digested with the restriction endonucleases BamH I and Xba I resulting in the removal nucleotides #1-#34 of LINK-1 (refer to the numbering of oligo #5). Clone ML14a, which contains an insert comprising the sequence set forth in SEQ ID NO:1, is also digested with the restriction endonucleases BamH I and Xba I resulting in the removal of sequences comprising nucloetides #1-#1515 of SEQUENCE ID NO:1 (murine BMP-9). This BamH I/Xba I fragment of mouse BMP-9 is isolated from the remainder of the ML14a plasmid clone and subcloned into the BamH I/Xba I sites generated by the removal of the synthetic linker sequences described above. The resulting clone is designated p302.

The p302 clone is digested with the restriction endonuclease EcoOlO9 I resulting in the excision of nucloetides corresponding to nucleotides #621-#1515 of the murine BMP-9

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sequence (SEQ ID NO:1) and nucleotides #35-#59 of LINK-1 (refer to numbering of oligonucleotide #5). It should be noted that the Apa I restriction site created in LINK-1 by the A to G conversion described above is a subset of the recognition sequence of EcoOlO9 I, therefore digestion of p302 with EcoOlO9 I cleaves at the Apa I site as well as the naturally occuring murine EcoOlO9 I (location #619-#625 of SEQ ID NO:1) resulting in the excision of a 920 bp EcoOlO9 I/EcoOlO9 I (Apa I) fragment comprising the sequences described above. This 920 EcoOlO9 I/EcoOlO9 I (Apa I) fragment is isolated from the remainder of the p302 plasmid clone and subcloned into clone pBMP-9link which has been similarly digested with EcoOl09 I. It should be noted that the nucleotides GG (#32-#33 of oligonucleotide #5) originally designed to facilitate a more complete digestion of the two adjacent restriction sites EcoO109 I and Xba I of LINK-1, which is now a part of pBMP-9link (described above), results in the creation of Dcm methylation recognition sequence. The restriction nuclease EcoOlO9 I is sensitive to Dcm methylation and therefore (nucleotides #25-#31 of cleavage of this sequence oligonucleotide #5/LINK-1) by the restriction endonuclease EcoOlO9 I is prevented at this site. Therefore the plasmid clone pBMP-9link is cleaved at the Apa I site but not at the EcoOlO9 I site upon digestion with the restriction endonuclease EcoOlO9 I as described above, preventing the intended removal of the sequences between the EcoOlO9 I and Xba I site of LINK-1

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(#32-#55 defined by the numbering of oligonucleotide #5). This results in the insertion of the 920 bp EcoOlO9 I/Apa I fragment at the EcoOlO9 I (Apa I) site of pBMP-9link. The resulting clone is designated p318.

Clone p318 is digested with the restriction endonucleases Sal I and Apa I, resulting in the excision of sequences comprising nucleotides #6-#56 of LINK-1 (refer to oligo #5 for location), nucleotides #621-#1515 of murine BMP-9 (SEQ ID NO:1), and nucleotides #35-#60 of LINK-1 (refer to oligo #5 for location). The resulting 972 bp Sal I/Apa I fragment described above is isolated from the remainder of the p318 plasmid clone and will be utilized in subsequent manipulations.

The clone phBMP9mex-1 (described above), which contains DNA sequences which—encode the entire mature region and portions of the propeptide of the human BMP-9 protein, is digested with the restriction endonucleases Apa I and EcoR I. This results in the excision of a 374 bp fragment comprising nucleotides #105-#470 of the human BMP-9 sequence (SEQ ID NO:8) and the additional nucleotides of oligonucleotide primers #3 and #4 which contain the recognition sequences for the restriction endonucleases Apa I and EcoR I. This 374 bp Apa I/EcoR I fragment is combined with the 972 bp Sal I/Apa I fragment from p138 (isolation described above) and ligated to the mammalian cell expression plasmid pED6 (a derivative of pEMC2β1) which has been digested with Sal I and EcoR I. The resulting clone is designated p324.

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The clone ML14a (murine BMP-9) is digested with EcoO109 I and Xba I to generate a fragment comprising nucleotides #621-#1515 of SEQ ID NO:1.

The following oligonucleotides are synthesized on an automated DNA synthesizer and combined such that their complimentary sequences can base pair (anneal) with each other to generate a double stranded synthetic DNA linker designated LINK-2:

#7 TCGACCACCATGTCCCCTGG

10 #8 GCCCCAGGGGACATGGTGG

This double stranded synthetic DNA linker (LINK-2) anneals in such a way that it generates single stranded ends which are compatible to DNA fragments digested with Sal I (one end) or EcoOlO9 I (the other end) as indicated below:

15 #7 TCGACCACCATGTCCCCTGG GGTGGTACAGGGGACCCCG #8.

This LINK-2 synthetic DNA linker is ligated to the 895 bp EcoOl09 I/Xba I fragment comprising nucleotides #621-#1515 of murine BMP-9 (SEQ ID NO:1) described above. This results in a 915 bp Sal I/Xba I fragment.

The clone p324 is digested with Sal I/Xba I to remove sequences comprising nucleotides #6-#56 of LINK-1 (refer to oligo #5 for location) and nucleotides #621-#1515 of murine BMP-9 (SEQ ID NO:1). The sequences comprising nucleotides #35-#60 of LINK-1 (refer to oligo #5 for location) and the sequences comprising the 374 bp Apa I/EcoR I fragment (human BMP-9 sequences) derived from phBMP9mex-1 remain attached to

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the pED6 backbone. The 915 bp Sal I/Xba I fragment comprising LINK-2 sequences and nucleotides #621-#1515 of murine BMP-9 (SEQ ID NO:1) is ligated into the p324 clone from which the Sal I to Xba I sequences described above have been removed.

The resulting plasmid is designated BMP9fusion and comprises LINK-2, nucleotides #621-#1551 of murine BMP-9 (SEQ ID NO:1), nucleotides #35-#59 of LINK-1 (refer to the numbering of oligonucleotide #5), and the 374 bp Apa I/EcoR I fragment (human BMP-9) derived from clone pBMP9mex-1 (described above) inserted between the Sal I and EcoR I sites of the mammalian cell expression vector pED6.

BMP9 fusion is transfected into CHO cells using standard techniques known to those having ordinary skill in the art to create—stable cell lines capable of expressing human BMP-9 protein. The cell lines are cultured under suitable culture conditions and the BMP-9 protein is isolated and purified from the culture medium.

EXAMPLE V

20 <u>Biological Activity of Expressed BMP-9</u>

To measure the biological activity of the expressed BMP-9 proteins obtained in Example IV above, the proteins are recovered from the cell culture and purified by isolating the BMP-9 proteins from other proteinaceous materials with which they are co-produced as well as from other contaminants. The purified protein may be assayed in accordance with the rat bone

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formation assay described in Example III.

Purification is carried out using standard techniques known to those skilled in the art. It is contemplated, as with other BMP proteins, that purification may include the use of Heparin sephanose.

Protein analysis is conducted using standard techniques such as SDS-PAGE acrylamide [U.K. Laemmli, Nature 227:680 (1970)] stained with silver [R.R. Oakley, et al. Anal. Biochem. 105:361 (1980)] and by immunoblot [H. Towbin, et al. Proc. Natl. Acad. Sci. USA 76:4350 (1979)]

The foregoing descriptions detail presently preferred embodiments of the present invention. Numerous modifications and variations in practice thereof are expected to occur to those skilled in the art upon consideration of these descriptions. Those modifications and variations are believed to be encompassed within the claims appended hereto.

- (1) GENERAL INFORMATION:
 - (i) APPLICANT: Wozney, John M. Celeste, Anthony
 - (ii) TITLE OF INVENTION: BMP-9 COMPOSITIONS
 - (iii) NUMBER OF SEQUENCES: 9
 - (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Genetics Institute, Inc.
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 - (C) CITY: Cambridge
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 - (E) COUNTRY: US
 - (F) ZIP: 02140 ...
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
 - (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: US
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
 - (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Kapinos, Ellen J.
 - (B) REGISTRATION NUMBER: 32,245
 - (C) REFERENCE/DOCKET NUMBER: GI 5186A
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: (617) 876-1170
 - (B) TELEFAX: (617) 876-5851
 - (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2447 base pairs

 - (B) TYPE: nucleic acid (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA to mRNA
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Mus musculus
 - (B) STRAIN: C57B46xCBA
 - (F) TISSUE TYPE: liver

(vii)	IMMEDIATE	SOURCE:
	THICHIAID	

(A) LIBRARY: Mouse liver cDNA (B) CLONE: ML14A

(viii) POSITION IN GENOME:

(C) UNITS: bp

(ix) FEATURE:

(A) NAME/KEY: mat_peptide
(B) LOCATION: 1564..1893

(ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 610..1896

(ix) FEATURE:

(A) NAME/KEY: mRNA

(B) LOCATION: 1..2447

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CATTAATAAA S	TATTAAGTAT	TGGAATTAGI	GAAATTGGA	G TTCCTTGTG	G AAGGAAGTGG	60				
GCAAGTGAGC	PTTTTAGTTT	GTGTCGGAAG	CCTGTAATT	A CGGCTCCAG	C TCATAGTGGA	120				
ATGGCTATAC :	TAGATTTAT	GGATAGTTGG	GTAGTAGGT	G TAAATGTAT	G TGGTAAAAGG	180				
CCTAGGAGAT	TGTTGATCC	AATAAATATG	ATTAGGGAA	A CAATTATTA	G GGTTCATGTT	240				
CGTCCTTTTG (STGTGTGGAT	TAGCATTATI	TGTTTGATA	A TAAGTTTAA	C TAGTCAGTGT	300				
TGGAAAGAAT (GAGACGGTT	GTTGATTAGG	CGTTTTGAG	G ATGGGAATA	G GATTGAAGGA	360				
AATATAATGA	rggctacaac	GATTGGGAAT	CCTATTATI	G TTGGGGTAA	r gaatgaggca	420				
AATAGATTTT (CGTTCATTTT	AATTCTCAAG	GGGTTTTTA	C TTTTATGTT	T GTTAGTGATA	480				
TTGGTGAGTA (GCCAAGGGT	TAATAGTGTA	ATTGAATTA	T AGTGAAATC	A TATTACTAGA	540				
CCTGATGTTA (GAAGGAGGGC	TGAAAAGGCI	CCTTCCCTC	C CAGGACAAA	A CCGGAGCAGG	600				
Mo	GCCACCCGG ATG TCC CCT GGG GCC TTC CGG GTG GCC CTG CTC CCG CTG Met Ser Pro Gly Ala Phe Arg Val Ala Leu Leu Pro Leu 648									
•	318	-315		-310						
TTC CTG CTG Phe Leu Leu -305	Val Cys V		Gln Lys Pr			696				
011 001 mag			010 100 m	00 000 001 M	TO MOTE CC.	744				
CAA GCA TCC Gln Ala Ser						/44				
GCT GGA GAG Ala Gly Glu	Glu Gly V		Leu Gln Me	et Phe Leu G	lu Asn Met	792				
_	-270		-265	-	260					
AAG GTG GAT Lys Val Asp						840				

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 151 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:
- * Thr Arg Glu Cys Ser Arg Ser Cys Pro Arg Thr Ala Pro Gln Arg
 -41 -40 -35 -30
- Gln Val Arg Ala Val Thr Arg Arg Thr Arg Met Ala His Val Ala Ala -25 -15 -10
- Gly Ser Thr Leu Ala Arg Arg Lys Arg Ser Ala Gly Ala Gly Ser His
- Cys Gln Lys Thr Ser Leu Arg Val Asn Phe Glu Asp Ile Gly Trp Asp
- Ser Trp Ile Ile Ala Pro Lys Glu Tyr Glu Ala Tyr Glu Cys Lys Gly
 25 30 35
- Gly Cys Phe Phe Pro Leu Ala Asp Asp Val Thr Pro Thr Lys His Ala 40 45 50 55
- Ile Val Gln Thr Leu Val His Leu Lys Phe Pro Thr Lys Val Gly Lys
 60 65 70
- Ala Cys Cys Val Pro Thr Lys Leu Ser Pro Ile Ser Val Leu Tyr Lys
 75 80 85
- Asp Asp Met Gly Val Pro Thr Leu Lys Tyr His Tyr Glu Gly Met Ser 90 95 100
- Val Ala Glu Cys Gly Cys Arg 105 110

(ix)	(A)	URE: NAM LOC	\mathbf{E}/\mathbf{KE}	Y: e N: 1	xon 47	0									
(ix)	(A)	URE: NAM LOC	E/KE	Y: C	DS	6									
(ix)	(A)	TURE: NAM LOC	压/KE	: И: : Т	at_p 24	epti 453	ide								
((ix)	(A	TURE:) NAI) LO	Æ/KI	EY: I	nRNA L47	70		•							
	(zi)	SEO	UENC:	E DES	SCRI	PTIO1	1: S	EQ I	D NO	:8:						
	ACA . Thr				TCA Ser			mem	CCA Pro	AGG Arg	ACG Thr -30	GCT (Ala (CCA Pro	CAG Gln	AGG Arg	48
CAG Gln -25		AGA Arg	GCA Ala	GTC Val	ACG Thr -20	AGG . Arg .	AGG Arg	ACA Thr	CGG Arg	ATG Met -15	GCG Ala	CAC His	GTG Val	GCT Ala	GCG Ala -10	96
GGG Gly	TCG Ser	ACT Thr	TTA Leu	GCC Ala -5	AGG Arg	Arg	пÀР	AGG Arg	AGC Ser	GCC Ala	GGG Gly	GCT Ala	GGC Gly . 5	AGC Ser	CAC His	144
TGT Cys	CAA Gln	AAG Lys 10	ACC Thr	TCC Ser	CTG Leu	CGG Arg	GTA Val 15	AAC Asn	TTC Phe	GAG Glu	GAC Asp	ATC Ile 20	GGC Gly	TGG Trp	GAC Asp	192
AGC Ser	TGG Trp 25	ATC Ile	ATT Ile	GCA Ala	CCC Pro	AAG Lys 30	GAG Glu	TAT Tyr	GAA Glu	GCC Ala	TAC Tyr 35	GAG Glu	TGT Cys	AAG Lys	GGC	240
GGC Gly	TGC Cys	TTC Phe	Phe	Pro	TTG Leu - 45	ATA	GAC Asp	GAT Asp	GTG Val	ACG Thr 50		ACG Thr	AAA Lys	CAC His	GCT Ala 55	288
		CAG Gln	ACC Thr	CTG Leu 60	val	CAT His	CTC Leu	AAG Lys	TTC Phe 65		ACA Thr	AAG Lys	GTG Val	GGC Gly 70	AAG Lys	336
GCC Ala	TGC	TGT	GTG Val	. Pro	ACC Thr	AAA Lys	CTG Lev	AGC Ser 80		ATC Ile	TCC Ser	GTC Val	CTC Lev	TAC Tyr	AAG Lys	384
GAT Asp	GAC Asp	ATO Mei	GGG Gly	•	CCC Pro	ACC Thr	CTC Lev	٠ ړت ٠	G TAC S Tyl	CAT His	TAC Tyl	GAG Glu	GGC Gly	ATO	G AGC Ser	432
GT0 Val	GCA Ala	a Gl	G TG! u Cy:	r GGC s Gly	TGC Y Cys	AGG Arg	3	GTAT(CTGC	CTG	CGGG					470

CATO	~~~	0300	3 mc	222
צוים")	-1-1-	(AL-I		T-AI-

-WIGGCWG	ic Tegas	
(2) INFOR	MATION FOR SEQ ID NO:6:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 34 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: cDNA to mRNA	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:6:	
CTGCAGGC	GA GCCTGAATTC CTCGAGCCAT CATG	34
(2) INFOR	RMATION FOR SEQ ID NO:7:	
(1)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 68 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: cDNA to mRNA	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:7:	
CGAGGTTA	AA AAACGTCTAG GCCCCCGAA CCACGGGGAC GTGGTTTTCC TTTGAAAAAC	60
ACGATTGC		68
(2) INFO	RMATION FOR SEQ ID NO:8:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 470 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: DNA (genomic)	
(iii)	HYPOTHETICAL: NO	
(v)	FRAGMENT TYPE: C-terminal	
(vi)	ORIGINAL SOURCE: (A) ORGANISM: Homo sapiens (H) CELL LINE: W138 (genomic DNA)	
(vii)	IMMEDIATE SOURCE: (A) LIBRARY: human genomic library (B) CLONE: lambda lll-l	

(viii) POSITION IN GENOME: (C) UNITS: bp

Arg Ile Asn Ile Tyr Glu Val Met Lys Pro Pro Ala Glu Val Val Pro -115 -105

Gly His Leu Ile Thr Arg Leu Leu Asp Thr Arg Leu Val His His Asn -100 -95 -90 -85

Val Thr Arg Trp Glu Thr Phe Asp Val Ser Pro Ala Val Leu Arg Trp
-80 -75 -70

Thr Arg Glu Lys Gln Pro Asn Tyr Gly Leu Ala Ile Glu Val Thr His

Leu His Gln Thr Arg Thr His Gln Gly Gln His Val Arg Ile Ser Arg
-50 -45 -40

Ser Leu Pro Gln Gly Ser Gly Asn Trp Ala Gln Leu Arg Pro Leu Leu
-35 -30 -25

Val Thr Phe Gly His Asp Gly Arg Gly His Ala Leu Thr Arg Arg Arg -20 -15 -10 -5

Arg Ala Lys Arg Ser Pro Lys His His Ser Gln Arg Ala Arg Lys Lys 1 5 10

Asn Lys Asn Cys Arg Arg His Ser Leu Tyr Val Asp Phe Ser Asp Val

Gly Trp Asn Asp Trp Ile Val Ala Pro Pro Gly Tyr Gln Ala Phe Tyr 30 35 40

Cys His Gly Asp Cys Pro Phe Pro Leu Ala Asp His Leu Asn Ser Thr
45 50 55 60

Asn His Ala Ile Val Gln Thr Leu Val Asn Ser Val Asn Ser Ser Ile
65 70 75

Pro Lys Ala Cys Cys Val Pro Thr Glu Leu Ser Ala Ile Ser Met Leu 80 85 90

Tyr Leu Asp Glu Tyr Asp Lys Val Val Leu Lys Asn Tyr Gln Glu Met 95 100 105

Val Val Glu Gly Cys Gly Cys Arg

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA to mRNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

TGT GGG TGC CGC TGAGATCAGG CAGTCC Cys Gly Cys Arg 115	CTTGA GGATAGACAG ATATACACAC	1666
CACACACACA CACCACATAC ACCACACACA	CACGTTCCCA TCCACTCACC CACACACTAC	1726
ACAGACTGCT TCCTTATAGC TGGACTTTTA	TTTAAAAAA AAAAAAAAA AATGGAAAAA	1786
ATCCCTAAAC ATTCACCTTG ACCTTATTTA	TGACTTTACG TGCAAATGTT TTGACCATAT	1846
TGATCATATA TTTTGACAAA ATATATTTAT	AACTACGTAT TAAAAGAAAA AAATAAAATG	1906
AGTCATTATT TTAAAAAAA AAAAAAACT	CTAGAGTCGA CGGAATTC	1954

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 408 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Ile Pro Gly Asn Arg Met Leu Met Val Val Leu Leu Cys Gln Val -292 -285 -280

Leu Leu Gly Gly Ala Ser His Ala Ser Leu Ile Pro Glu Thr Gly Lys
-275 -270 -265

Lys Lys Val Ala Glu Ile Gln Gly His Ala Gly Gly Arg Arg Ser Gly -260 -255 -250 -245

Gln Ser His Glu Leu Leu Arg Asp Phe Glu Ala Thr Leu Leu Gln Met
-240 -235 -230

Phe Gly Leu Arg Arg Pro Gln Pro Ser Lys Ser Ala Val Ile Pro
-225 -220 -215

Asp Tyr Met Arg Asp Leu Tyr Arg Leu Gln Ser Gly Glu Glu Glu Glu -210 -205 -200

Glu Gln Ile His Ser Thr Gly Leu Glu Tyr Pro Glu Arg Pro Ala Ser
-195 -190 -185

Arg Ala Asn Thr Val Arg Ser Phe His His Glu Glu His Leu Glu Asn -180 -175 -170 -165

Ile Pro Gly Thr Ser Glu Asn Ser Ala Phe Arg Phe Leu Phe Asn Leu
-160 -155 -150

Ser Ser Ile Pro Glu Asn Glu Val Ile Ser Ser Ala Glu Leu Arg Leu
-145 -140 -135

Phe Arg Glu Gln Val Asp Gln Gly Pro Asp Trp Glu Arg Gly Phe His
-130 -125 -120

				Pro					Gly			CGT Arg		Asn		942
TAT Tyr	GAG Glu	GTT Val -110	Met	AAG Lys	CCC Pro	CCĀ Prọ	GCA Ala -105	Glu	GTG Val	GTG Val	CCT Pro	GGG Gly -100	His	CTC Leu	ATC Ile	990
												GTG Val			TGG Trp	1038
												ACC Thr				1086
CAG Gln	CCA Pro	AAC Asn	TAT Tyr	GGG Gly -60	CTA Leu	GCC Ala	ATT Ile	GAG Glu	GTG Val -55	ACT Thr	CAC His	CTC Leu	CAT His	CAG Gln -50	ACT Thr	1134
CGG Arg	ACC Thr	CAC His	CAG Gln -45	GGC Gly	CAG Gln	CAT His	GTC Val	AGG Arg -40	ATT Ile	AGC Ser	CGA Arg	TCG Ser	TTA Leu -35	CCT Pro	CAA Gln	1182
GGG Gly	AGT Ser	GGG Gly -30	AAT Asn	TGG Trp	GCC Ala	CAG	CTC Leu -25	CĠG Arg	ccċ Pro	CTC Leu	ČŤG Leu	GTĆ Val -20	ÁCC Thr	TTT Phe	GGC	1230
					His							AGG Arg				1278
												AAT Asn				1326
CGG	CGC Arg	CAC His	TCG Ser 20	CTC Leu	TAT Tyr	GTG Val	GAC Asp	TTC Phe 25	AGC Ser	GAT Asp	GTG Val	GGC Gly	TGG Trp 30	AAT Asn	-GAC Asp	1374
TGG Trp	ATT Ile	GTG Val 35	GCC Ala	CCA Pro	CCA Pro	GGC Gly	TAC Tyr 40	CAG Gln	GCC Ala	TTC Phe	TAC Tyr	TGC Cys 45	CAT His	GGG Gly	GAC Asp	1422
TGC Cys	CCC Pro 50	Phe	CCA Pro	CTG Leu	GCT Ala	GAC Asp 55	CAC His	CTC Leu	AAC Asn	TCA Ser	ACC Thr 60	AAC Asn	CAT His	GCC Ala	ATT Ile	1470
GTG Val 65	CAG Gln	ACC Thr	CTG Leu	GTC Val	AAT Asn 70	TCT Ser	GTC Val	AAT Asn	TCC Ser	AGT Ser 75	ATC Ile	CCC Pro	AAA Lys	GCC Ala	TGT Cys 80	1518
TGT Cys	GTG Val	CCC	ACT Thr	GAA Glu 85	Leu	AGT Ser	GCC Ala	ATC Ile	TCC Ser 90	ATG Met	CTG Leu	TAC Tyr	CTG Leu	GAT Asp 95	GAG Glu	1566
TAT Tyr	GAT Asp	AAG Lys	GTG Val 100	GTA Val	CTG Leu	AAA Lys	AAT Asn	TAT Tyr 105	CAG Gln	GAG Glu	ATG Met	GTA Val	GTA Val 110	GAG Glu	GGA Gly	1614

WO 93/00432 PCT/US92/05374

(B) LOCATION: 9..1934

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

	CTCT	AGAG	GG C	AGAG	GAGG	A GG	GAGG	GAGG	GAA	GGAG	CGC	GGAG	CCCG	GC C	CGGA	AGCTA		60
	GGTG	AGTG	TG G	CATC	CGAG	C TG	AGGG	ACGC	GAG	CCTG	AGA	CGCC	GCTG	CT G	CTCC	GGCTG		120
	AGTA	TCTA	GC T	TGTC	TCCC	C GA	TGGG	ATTC	CCG	TCCA	AGC	TATC	TCGA	GC C	TGCA	GCGCC		180
	ACAG	TCCC	CG G	CCCT	CGCC	C AG	GTTC	ACTG	CAA	CCGT	TCA	GAGG	TCCC	CA G	GAGC	TGCTG		240
	CTGG	CGAG	cc c	GCTA	CTGC	A GG	GACC	TATG	GAG	CCAT	TCC	GTAG	TGCC	AT C	CCGA	GCAAC		300
	GCAC	TGCT	GC A	GCTT.	CCCT	G AG	CCTT	TCCA	GCA	AGTT	TGT	TCAA	GATT	GG C	TGTC	AAGAA		360
	TCAT	'GGAC	TG T	TTATT	ATAT	G CC	TTGT	TTTC	TGT	CAAG	ACA	M	TG A let I 292	le P				414
				CTG Leu -285	Met					Cys			Leu		Gly			462
				GCT Ala					Glu					Lys				510
*: :-			Gln					Gly					Gln			GAG Glu	-	558
		Leu		GAC Asp			Ala					Met						606
				CAG Gln		Ser					Ile							654
				CGG Arg -205	Leu					Glu					Ile	CAC His		702
				CTT Leu O					Arg					Ala				750
			Ser	TTC Phe				Glu					Ile					798
		Glu		TCT Ser			Arg					Leu						846
				GTG Val		Ser					Arg					Gln		894

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Gly Ala Ser Ser His Cys Gln Lys Thr Ser Leu Arg Val Asn Phe Glu

Asp Ile Gly Trp Asp Ser Trp Ile Ile Ala Pro Lys Glu Tyr Asp Ala

Tyr Glu Cys Lys Gly Gly Cys Phe Phe Pro Leu Ala Asp Asp Val Thr

Pro Thr Lys His Ala Ile Val Gln Thr Leu Val His Leu Glu Phe Pro

Thr Lys Val Gly Lys Ala Cys Cys Val Pro Thr Lys Leu Ser Pro Ile

Ser Ile Leu Tyr Lys Asp Asp Met Gly Val Pro Thr Leu Lys Tyr His

Tyr Glu Gly Met Ser Val Ala Glu Cys Gly Cys Arg 105 100

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1954 base pairs (B) TYPE: nucleic acid

 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- ---- (ii) MOLECULE TYPE: cDNA to mRNA
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Homo sapiens
 - (G) CELL TYPE: Osteosarcoma Cell Line
 - (H) CELL LINE: U-20S
 - (vii) IMMEDIATE SOURCE:
 - (A) LIBRARY: U2OS cDNA in Lambda gt10
 - (B) CLONE: Lambda U2OS-3
 - (viii) POSITION IN GENOME:
 - (C) UNITS: bp
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 403..1629
 - (ix) FEATURE:
 - (A) NAME/KEY: mat_peptide
 - (B) LOCATION: 1279...1626
 - (ix) FEATURE:
 - (A) NAME/KEY: mRNA

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:
- Met Ser Pro-Gly Ala Phe Arg Val Ala Leu Leu Pro Leu Phe Leu Leu -318 -315 -305
- Val Cys Val Thr Gln Gln Lys Pro Leu Gln Asn Trp Glu Gln Ala Ser
 -300 -295 -290
- Pro Gly Glu Asn Ala His Ser Ser Leu Gly Leu Ser Gly Ala Gly Glu
 -285 -280 -275
- Glu Gly Val Phe Asp Leu Gln Met Phe Leu Glu Asn Met Lys Val Asp
 -270 -265 -260 -255
- Phe Leu Arg Ser Leu Asn Leu Ser Gly Ile Pro Ser Gln Asp Lys Thr
 -250 -245 -240
- Arg Ala Glu Pro Pro Gln Tyr Met Ile Asp Leu Tyr Asn Arg Tyr Thr
 -235 -230 -225
- Thr Asp Lys Ser Ser Thr Pro Ala Ser Asn Ile Val Arg Ser Phe Ser -220 -215 -210
- Val Glu Asp Ala Ile Ser Thr Ala Ala Thr Glu Asp Phe Pro Phe Gln
 -205 -200 -195
- Lys His Ile Leu Ile Phe Asn Ile Ser Ile Pro Arg His Glu Gln Ile
 -190 -185 -180 -175
- Thr Arg Ala Glu Leu Arg Leu Tyr Val Ser Cys Gln Asn Asp Val Asp
 -170 -165 -160
- Ser Thr His Gly Leu Glu Gly Ser Met Val Val Tyr Asp Val Leu Glu
 -155 -150 -145
- Asp Ser Glu Thr Trp Asp Gln Ala Thr Gly Thr Lys Thr Phe Leu Val -140 -135 -130
- Ser Gln Asp Ile Arg Asp Glu Gly Trp Glu Thr Leu Glu Val Ser Ser -125 -120 -115
- Ala Val Lys Arg Trp Val Arg Ala Asp Ser Thr Thr Asn Lys Asn Lys
- Leu Glu Val Thr Val Gln Ser His Arg Glu Ser Cys Asp Thr Leu Asp
- Ile Ser Val Pro Pro Gly Ser Lys Asn Leu Pro Phe Phe Val Val Phe
 -75 -70 -65
- Ser Asn Asp Arg Ser Asn Gly Thr Lys Glu Thr Arg Leu Glu Leu Lys
 -60 -55 -50
- Glu Met Ile Gly His Glu Gln Glu Thr Met Leu Val Lys Thr Ala Lys
 -45 -40 -35
- Asn Ala Tyr Gln Val Ala Gly Glu Ser Gln Glu Glu Glu Gly Leu Asp
 -30 -25 -20 -15
- Gly Tyr Thr Ala Val Gly Pro Leu Leu Ala Arg Arg Lys Arg Ser Thr

- 15	-10	-5	
AGG AGC ACC GGA GC Arg Ser Thr Gly Al	C AGC AGC CAC TGC a Ser Ser His Cys 5	CAG AAG ACT TCT Gln Lys Thr Ser 10	CTC AGG GTG 1608 Leu Arg Val 15
AAC TTT GAG GAC AT Asn Phe Glu Asp Il 2	e Gly Trp Asp Ser	TGG ATC ATT GCA Trp Ile Ile Ala 25	CCC AAG GAA 1656 Pro Lys Glu 30
TAT GAC GCC TAT GA Tyr Asp Ala Tyr Gl 35	G TGT AAA GGG GGT u Cys Lys Gly Gly 40	Cys Phe Phe Pro	TTG GCT GAT 1704 Leu Ala Asp 45
GAC GTG ACA CCC AC Asp Val Thr Pro Th	C AAA CAT GCC ATC r Lys His Ala Ile 55	GTG CAG ACC CTG Val Gln Thr Leu 60	GTG CAT CTC 1752 Val His Leu
GAG TTC CCC ACA AA Glu Phe Pro Thr Ly 65	G GTG GGC AAA GCC s Val Gly Lys Ala 70	TGC TGC GTT CCC Cys Cys Val Pro 75	ACC AAA CTG 1800 Thr Lys Leu
AGT CCC ATC TCC AT Ser Pro Ile Ser Il 80	C CTC TAC AAG GAT e Leu Tyr Lys Asp 85	GAC ATG GGG GTG Asp Met Gly Val 90	CCA ACC CTC 1848 Pro Thr Leu 95
AAG TAC CAC TAT GA Lys Tyr His Tyr Gl	u Gly Met Ser Val	GCT GAG TGT GGG Ala Glu Cys Gly 105	TGT AGG TAGTCCCTGC 1903 Cys Arg 110
AGCCACCCAG-GGTGGGG	ATA-CAGGACATGG AA	GAGGTTCT GGTACGG	TCC TGCATCCTCC 1963
TGCGCATGGT ATGCCTA	AGT TGATCAGAAA CC	ATCCTTGA GAAGAAA	AGG AGTTAGTTGC 2023
CCTTCTTGTG TCTGGTG	GGT CCCTCTGCTG AA	GTGACAAT GACTGGG	GTA TGCGGGCCTG 2083
TGGGCAGAGC AGGAGAC	CCT GGAAGGGTTA GT	GGGTAGAA AGATGTC	AAA AAGGAAGCTG 2143
TGGGTAGATG ACCTGCA	CTC CAGTGATTAG AA	GTCCAGCC TTACCTG	TGA GAGAGCTCCT 2203
GGCATCTAAG AGAACTC	TGC TTCCTCATCA TC	CCCACCGA CTTGTTC	TTC CTTGGGAGTG 2263
TGTCCTCAGG GAGAACA	GCA TTGCTGTTCC TG	TGCCTCAA GCTCCCA	GCT GACTCTCCTG 2323
TGGCTCATAG GACTGAA	TGG GGTGAGGAAG AG	CCTGATGC CCTCTGG	CAA TCAGAGCCCG 2383
AAGGACTTCA AAACATC	TGG ACAACTCTCA TT	GACTGATG CTCCAAC	ATA ATTTTTAAAA 2443
			2447

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 428 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

-255 -250 -245

233		200			
GAC AAA ACC AGA Asp Lys Thr Arg -240	GCG GAG CCA Ala Glu Pro -235	Pro Gln Tyr	ATG ATC GAC T Met Ile Asp L -230	TG TAC AAC eu Tyr Asn	888
AGA TAC ACA ACG Arg Tyr Thr Thr -225	GAC AAA TCG Asp Lys Ser -220	TCT ACG CCT Ser Thr Pro	GCC TCC AAC A Ala Ser Asn I -215	TC GTG CGG le Val Arg -210	936
AGC TTC AGC GTG Ser Phe Ser Val			Ala Ala Thr G		984
CCC TTT CAG AAC Pro Phe Gln Lys	His Ile Leu	ATC TTC AAC Ile Phe Asn -185	Ile Ser Ile P	CG AGG CAC ro Arg His 180	1032
GAG CAG ATC ACC Glu Gln Ile Thi -175	AGG GCT GAG Arg Ala Glu	CTC CGA CTC Leu Arg Leu -170	TAT GTC TCC T Tyr Val Ser C -165	GC CAA AAT Ys Gln Asn	1080
GAT GTG GAC TCC Asp Val Asp Sec -160	C ACT CAT GGG T Thr His Gly -159	Leu Glu Gly	AGC ATG GTC G Ser Met Val V -150	TT TAT GAT al Tyr Asp	1128
GTT CTG GAG GAG Val Leu Glu Asp -145	C AGT GAG ACT p Ser Glu Thr -140	TGG GAC CAG Trp Asp Gln	GCC ACG GGG A Ala Thr Gly T -135	CC AAG ACC Thr Lys Thr -130	1176
TTC TTG GTA TC	r Gln Asp Ile -125	Arg Asp Glu	o Gly Trp Glu I	Thr Leu Glu -115	1224
GTA TCG AGT GC Val Ser Ser Al -1	a Val Lys Arg	TGG GTC AGG Trp Val Arg -105	Ala Asp Ser ?	ACA ACA AAC Thr Thr Asn -100	1272
AAA AAT AAG CT Lys Asn Lys Le -95	C GAG GTG ACA u Glu Val Thr	GTG CAG AGG Val Gln Ser -90	C CAC AGG GAG A r His Arg Glu S -85	AGC TGT GAC Ser Cys Asp	1320
ACA CTG GAC AT Thr Leu Asp Il -80	C AGT GTC CCT e Ser Val Pro -75	Pro Gly Se	C AAA AAC CTG (r Lys Asn Leu) -70	CCC TTC TTT Pro Phe Phe	1368
GTT GTC TTC TC Val Val Phe Se -65	C AAT GAC CGC ar Asn Asp Arg -60	AGC AAT GG Ser Asn Gl	G ACC AAG GAG y Thr Lys Glu -55	ACC AGA CTG Thr Arg Leu -50	1416
GAG CTG AAG GA Glu Leu Lys Gl	AG ATG ATC GGC Lu Met Ile Gly -45	CAT GAG CA His Glu Gl -4	G GAG ACC ATG n Glu Thr Met 0	CTT GTG AAG Leu Val Lys -35	1464
Thr Ala Lys As	AT GCT TAC CAC sn Ala Tyr Glr 30	G GTG GCA GG n Val Ala Gl -25	T GAG AGC CAA y Glu Ser Gln	GAG GAG GAG Glu Glu Glu -20	1512
GGT CTA GAT GG Gly Leu Asp G	GA TAC ACA GCT Ly Tyr Thr Ala	GTG GGA CC A Val Gly Pr	A CTT TTA GCT o Leu Leu Ala	AGA AGG AAG Arg Arg Lys	1560

What is claimed is:

- 1. A BMP-9 polypeptide comprising the amino acid sequence from amino acid #8 110 as set forth in FIG. 3 (SEQ ID NO: 9).
- 2. A BMP-9 polypeptide comprising the amino acid sequence from amino acid #1 - 110 as set forth in FIG. 3 (SEQ ID NO: 9).
- 3. A BMP-9 polypeptide of claim 1 wherein said polypeptide is a dimer wherein each subunit comprises at least the amino acid sequence from amino acid #8 110 of FIG. 3 (SEQ ID NO: 9).
- 4. A BMP-9 polypeptide of claim 2 wherein said polypeptide is a dimer wherein each subunit comprises at least the amino acid sequence from amino acid #1-110 of FIG. 3. (SEQ ID NO: 9).
- 5. A purified BMP-9 protein produced by the steps of
- (a) culturing a cell transformed with a cDNA comprising the nucleotide sequence from nucleotide #124 to #453 as shown in FIG. 3 (SEQ ID NO: 8); and
- (b) recovering and purifying from said culture medium a protein comprising the amino acid sequence from amino acid #1 to amino acid #110 as shown in FIG. 3 (SEQ ID NO: 9).
- 6. A purified BMP-9 protein produced by the steps of
 - (a) culturing a cell transformed with a cDNA comprising

the nucleotide sequence from nucleotide #124 to #453 as shown in FIG. 3 (SEQ ID NO: 8); and

- (b) recovering form said culture medium a protein comprising an amino acid sequence from amino acid #8 to amino acid #110 as shown in Figure 3 (SEQ ID NO: 9).
- 7. A BMP-9 protein characterized by the ability to induce the formation of cartilage and/or bone.
- 8. A DNA sequence encoding a BMP-9 protein.
- 9. The DNA sequence of claim 8 wherein said DNA comprises
 - (a) nucleotide 124 to 453 (SEQ ID NO: 8); and
- (b) sequences which hybridize thereto under stringent hybridization conditions and exhibit the ability to form cartilage and/or bone.
- 10. The DNA sequence of claim 8 wherein said DNA comprises
 - (a) nucleotide 145 to 453 (SEQ ID NO: 8); and
- (b) sequences which hybridize thereto under stringent hybridization conditions and exhibit the ability to form cartilage and/or bone.
- 11. A host cell transformed with a DNA sequence encoding BMP-8.

- 12. A method for producing a purified BMP-9 protein said method comprising the steps of
- (a) culturing a cell transformed with a cDNA comprising the nucleotide sequence encoding a BMP-9 protein; and
- (b) recovering and purifying said BMP-9 protein from the culture medium.
- 13. A pharmaceutical composition comprising an effective amount of a BMP-9 protein in admixture with a pharmaceutically acceptable vehicle.
- 14. A composition of claim 13 further comprising a matrix for supporting said composition and providing a surface for bone and/or cartilage growth.
- 15. The composition of claim 14 wherein said matrix comprises a material selected from the group consisting of hydroxyapatite, collagen, polylactic acid and tricalcium phosphate.
- 16. A method for inducing bone and/or cartilage formation in a patient in need of same comprising administering to said patient an effective amount of the composition of claim 13.
- 17. A pharmaceutical composition for wound healing and tissue repair said composition comprising an effective amount of the

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protein of a BMP-9 protein in a pharmaceutically acceptable vehicle.

18. A method for treating wounds and/or tissue repair in a patient in need of same comprising administering to said patient an effective amount of the composition of claim 17.

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Figure 1A

			10		20			30			40		5			60			70
(CATT	\ATA!	AA TA	ATTA	GTAI	TGG	AATT	AGT	GAAA'	TTGG.	AG T	TCCT	rgtgo	AAC	GAAG	TGG	GCAA	GTGA	GC
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-		19	50	.6166	160)	GIAA	170	CGGC		80	CATA	19 19		GCTF	200			
C	GAT			'AGTA			ATGT		TGGT			CTAG			מארים:			2 2002	210 ~~
		22			230			240			50		26			270	win		280
7	ATTAC	GGAI	AA CA	ATTA	TTAG	GGT	TCAT	GTT	CGTC	CTTT	IG G	TGTGT	rggai	TAG	CATI		TGTT	TGAT	AA
		29	90		300)		310		3	20		33	0		340			350
3	[AAG]	LATT	AC TA	GTCA	GTGI	TGG	AAAG	AAT	GGAG			TTGAT	PTAGO	CGI	TTTC			GAAT.	AG
,	n n mm e	36		~~~	370			380			90		40			410		4	420
·	MIT	AAGC		TATA			CTAC		GATT(CTATI			GGGI		GAAT		
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•	MINC	50	וו כפ	TICE	510	WAT	TCTC	520	GGGT		AC T 30	TTTAT	1GTT1 54		AGTO		TIGG		
C	GCC			מידע			ייע ע כי		እርጥር			ATTAC			ነር እመረ	550	C		560
			570		58		Omi	59		ww.	600		60		GAIG		GAAG 18	GAGG	GC
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	TGA	AAGG	CT (CCTT	CCTC	cc cz	AGGA	CAAA	A CC	GAG	CAGG	GCC	ACCC	G A	rg T	व्ट व्य	CT G	G	
														M	S	P			
		627			636			645			654			663			672		
	GCC	TTC	CGG	GTG	GCC	CTIC	CTC		CTG	TTC	cmc	CTG	GTC			-			
•	A	F	R	V	A	L	L	P	L	F	L	L	V	TGT C	GTC V	ACA T	CAG		
		•	-`	•				•	ם	r	بر	ע	V	C	V	T	Q	Q	
		681			690			699			708			717			726		
	AAG				AAC	TGG	GAA	CAA	GCA	TCC	CCT	GGG	GAA	$\overline{\mathtt{AAT}}$	GCC	CAC	AGC	TCC	
	K	P	L	Q	N	W	E	Q	A	S.	P	G	E	N	A	H	S	S	
		725			744						:				•				
		735			744			753			762			771			780		
	CTG	GGA	TTG	TCT	GGA	GCT	GGA	<u> </u>	GAG	CCM	GTC	TTT	GAC	CTG	272	N MC	TTC		
	L	G	L	S	G	A	G	E	E	G	V	F	D	L	O CAG	M	F	L	
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		789			798			807			816			825			834		
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	E	N	M	ĸ	V	ח	돠	Т.	D	C	Т.	N	T	•	C	т	D	•	

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Figure 1B

	843			852			861			870			879			888	
CAG Q	GAC D	AAA K	ACC T	AGA R	GCG A	GAG E	CCA P	CCC P	CAG Q	TAC Y	ATG M	ATC I	GAC D	TTG L	TAC Y	AAC N	AGA R
	897			906			915			924			933			942	
TAC Y	ACA T	ACG T	GAC D	AAA K	TCG S	TCT S	ACG T	CCT P	GCC A	TCC S	AAC N	ATC I	GTG V	CGG R	AGC S	TTC F	AGC S
	951			960			969			978			987			996	
GTG V	GAA E	GAT D	GCT A	ATA I	TCG S	ACA T	GCT A	GCC A	ACG T	GAG E	GAC D	TTC F	CCC P	TTT F	CAG Q	AAG K	CAC H
:	1005			1014		1	L023		:	1032		:	1041		:	1050	
ATC I	CTG L	ATC I	TTC F	AAC N	ATC I	TCC S	ATC I	CCG P	AGG R	CAC H	GAG E	CAG Q	ATC I	ACC T	AGG R	GCT A	GAG E
:	1059		:	1068		:	L077			1086		:	1095		:	1104	
CTC L	CGA R	CTC L	TAT Y	GTC V	TCC S	TGC C	CAA Q	AAT N	GAT D	GTG V	GAC D	TCC S	ACT T	CAT H	GGG G	CTG L	GAA E
:	1113		:	1122			131		:	1140		:	1149		:	1158	
<u> </u>																	
G	AGC S	ATG M	GTC V	GTT V	TAT Y	GAT D	GTT V	CTG L	GAG E	GAC D	AGT S	GAG E	ACT T	TGG W	GAC D	CAG Q	GCC A
G	AGC S 1167	ATG M	V	GTT V 1176	TAT Y	D			E			E			D		
G :	S 1167	M	v :	V 1176	Y	D :	V 1185	L	E	D 1194	S	E :	T 1203	W .	D :	Q	A
G ACG T	S 1167 GGG	M ACC	V AAG K	V 1176 ACC	Y TTC	D TTG L	V 1185 GTA	L TCC	E CAG Q	D 1194 GAC	S ATT	E CGG R	T 1203 GAC	<u>GAA</u>	D GGA G	Q 1212 TGG	A GAG
G ACG T	S 1167 GGG G 1221	M ACC T	V AAG K	V 1176 ACC T	Y TTC F	TTG L	V L185 GTA V L239	TCC S	E CAG Q	D 1194 GAC D 1248	S ATT I	E CGG R	T 1203 GAC D 1257	W GAA E	D GGA G	Q 1212 TGG W	GAG E
ACG T	S 1167 GGG G 1221 TTA	M ACC T	AAG K GTA V	V 1176 ACC T 1230 TCG	Y TTC F	TTG L GCC A	V L185 GTA V L239 GTG	TCC S	CAG Q CGG R	D 1194 GAC D 1248 TGG	S ATT I	E CGG R AGG R	T 1203 GAC D 1257 GCA	W GAA E	GGA G TCC S	Q 1212 TGG W 1266	GAG E
ACG T	S 1167 GGG G 1221 TTA L	M ACC T GAA E	V AAG K GTA V	V 1176 ACC T 1230 TCG S	TTC F	TTG L GCC A	V GTA V 1239 GTG V	TCC S AAG K	CAG Q CGG R	D 1194 GAC D 1248 TGG W	ATT I GTC V	E CGG R AGG R	T 1203 GAC D 1257 GCA A	GAA E GAC D	GGA G TCC S	Q 1212 TGG W 1266 ACA	GAG E ACA
ACG T ACT T	S 1167 GGG G 1221 TTA L 1275	M ACC T GAA E	AAG K GTA V	V 1176 ACC T 1230 TCG S 1284	TTC F AGT S	TTG L GCC A GTG V	V GTA V L239 GTG V L293 ACA	TCC S AAG K	CAG Q CAG Q	D 1194 GAC D 1248 TGG W	ATT I GTC V	E CGG R AGG R	T 1203 GAC D 1257 GCA A 1311	GAA E GAC D	GGA G TCC S	Q 1212 TGG W 1266 ACA T	GAG E ACA T

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Figure 1C

	1383	ı		1392	:		1401			1410			1419			1428	
TCC S	AAT N	GAC D	CGC R	AGC S	AAT N	GGG G	ACC T	AAG K	GAG E	ACC T	AGA R	CTC	GAG		AAG K		
	1437			1446			1455			1464			1473			1482	
ATC I	GGC G	CAT H	GAG E	CAG Q	GAG E	ACC T	ATG M	CTT L	GTG V	AAG K	ACA T	GCC A	AAA K	AAT N	GCT A	TAC Y	CAG Q
	1491			1500			1509			1518			1527			1536	
GTG V	GCA A	GGT G	GAG E	AGC S	CAA Q	GAG E	GAG E	GAG E	GGT G	CTA L	GAT D	GGA G	TAC	ACA T	GCT A	GTG V	GGA G
	1545			1554			1563	•		1572			1581			1590	
CCA P	L L	TTA L	GCT A	AGA R	AGG R	AAG K	1	AGC S 319)	ACC T	GGA G	GCC A	AGC S	AGC S	H	С	CAG Q	AAG K
	1599			1608			1617	•		1626			1635			1644	
ACT T	TCT S	CTC L	AGG R	GTG V	AAC N	TTT F	GAG E	GAC D	ATC I	GGC G	TGG W	GAC D	AGC S	TGG W	ATC I	ATT I	GCA A
	1653			1662			1671			1680			1689			L698	
P	AAG K	GAA E	TAT Y	GAC D	GCC A	TAT Y	GAG E	C		GGG G	GGT G	TGC C	TTC F	TTC F	CCA P		GCT A
•	1707			1716			L 72 5			L734			1743			752	
GAT D	GAC D	GTG V	ACA T	CCC P	ACC T	AAA K	CAT H	GCC A	ATC I	GTG V	CAG Q	ACC T	CTG L	GTG V	CAT H	CTC L	GAG E
	1761			1770			.779			1788			1797			.806	
TTC F	CCC P	ACA T	AAG K	GTG V	GGC G	AAA K	GCC A	TGC C	TGC C	GTT V	CCC P	ACC T	AAA K	CTG L	AGT S	CCC P	ATC I
	1815			L824			.833			.842			L851			860	
TCC S	ATC I	CTC L	TAC Y	AAG K	GAT D	GAC D	ATG M	GGG G	GTG V	CCA P	ACC T	CTC L	AAG K	TAC Y	CAC H	TAT Y	GAG E
	1869			1878			887					03		191			1923
GGG G	ATG M	AGT S	G⊒G ∵	GCT A	GAG E	TGT C	GGG G	C	AGG R 28)	TAGT	CCCT	GC A	GCCA	CCCA	.G GG	TGGG	gata

Figure 1D

1933	1943	1953			1983	1993
CAGGACATGG	AAGAGGTTCT	GGTACGGTCC	TGCATCCTCC	TGCGCATGGT	ATGCCTAAGT	TGATCAGAAA
2003	2013	2023	2033	2043	2053	2063
CCATCCTTGA	GAAGAAAAGG	AGTTAGTTGC	CCTTCTTGTG	TCTGGTGGGT	CCCTCTGCTG	AAGTGACAAT
2073	2083	2093	2103	2113	2123	2123
GACTGGGGTA	TGCGGGCCTG	TGGGCAGAGC	AGGAGACCCT	GGAAGGGTTA	GTGGGTAGAA	AGATGTCAAA
2143	2153	2163	2173	2183	2193	2203
AAGGAAGCTG	TGGGTAGATG	ACCTGCACTC	CAGTGATTAG	AAGTCCAGCC	TTACCTGTGA	GAGAGCTCCT
2213	2223	2233	2243	2253	2263	2273
GGCATCTAAG	AGAACTCTGC	TTCCTCATCA	TCCCCACCGA	CTTGTTCTTC	CTTGGGAGTG	TGTCCTCAGG
2283	2293	2303	2313	2323	2333	23/3
GAGAACAGCA	TTGCTGTTCC	TGTGCCTCAA	GCTCCCAGCT	GACTCTCCTG	TGGCTCATAG	GACTGAATGG
2353	2363	2373			2403	2413
GGTGAGGAAG	AGCCTGATGC	CCTCTGGCAA	TCAGAGCCCG	AAGGACTTCA	AAACATCTGG	ACAACTCTCA
242:	3 243	3 244	.3			3.2.2
TTGACTGAT	G CTCCAACAT.	A ATTTTTAAA	A AGAG			

518

Figure 2

10 20 30 40 50 60 70 CTCTAGAGGG CAGAGGAGGA GGGAGGGAGG GAAGGAGCGC GGAGCCCGGC CCGGAAGCTA GGTGAGTGTG
80 90 100 110 120 130 140 GCATCCGAGC TGAGGGACGC GAGCCTGAGA CGCCGCTGCT GCTCCGGCTG AGTATCTAGC TTGTCTCCCC
150 160 170 180 190 200 210 GATGGGATTC CCGTCCAAGC TATCTCGAGC CTGCAGCGCC ACAGTCCCCG GCCCTCGCCC AGGTTCACTG
220 230 240 250 260 270 280 CAACCGTTCA GAGGTCCCCA GGAGCTGCTG CTGGCGAGCC CGCTACTGCA GGGACCTATG GAGCCATTCC
290 300 310 320 330 340 350 GTAGTGCCAT CCCGAGCAAC GCACTGCTGC AGCTTCCCTG AGCCTTTCCA GCAAGTTTGT TCAAGATTGG
360 370 380 390 400 (1) CTGTCAAGAA TCATGGACTG TTATTATATG CCTTGTTTTC TGTCAAGACA CC ATG ATT CCT MET Ile Pro
417 432 447 462 GGT AAC CGA ATG CTG ATG GTC GTT TTA TTA TGC CAA GTC CTG CTA GGA GGC GCG Gly Asn Arg MET Leu MET Val Val Leu Leu Cys Gln Val Leu Leu Gly Gly Ala
477 492 507 AGC CAT GCT AGT TTG ATA CCT GAG ACG GGG AAG AAA AAA GTC GCC GAG ATT CAG Ser His Ala Ser Leu Ile Pro Glu Thr Gly Lys Lys Lys Val Ala Glu Ile Gln
522 567 GGC CAC GCG GGA GGA CGC CGC TCA GGG CAG AGC CAT GAG CTC CTG CGG GAC TTC Gly His Ala Gly Gly Arg Arg Ser Gly Gln Ser His Glu Leu Leu Arg Asp Phe
582 597 612 627 GAG GCG ACA CTT CTG CAG ATG TTT GGG CTG CGC CGC CCG CAG CCT AGC AAG Glu Ala Thr Leu Leu Gln MET Phe Gly Leu Arg Arg Pro Gln Pro Ser Lys
642 657 672 AGT GCC GTC ATT CCG GAC TAC ATG CGG GAT CTT TAC CGG CTT CAG TCT GGG GAG Ser Ala Val Ile Pro Asp Tyr MET Arg Asp Leu Tyr Arg Leu Gln Ser Gly Glu
687 702 717 732 GAG GAG GAA GAG CAG ATC CAC AGC ACT GGT CTT GAG TAT CCT GAG CGC CCG GCC Glu Glu Glu Gln Ile His Ser Thr Gly Leu Glu Tyr Pro Glu Arg Pro Ala
747 762 777 AGC CGG GCC AAC ACC GTG AGG AGC TTC CAC CAC GAA GAA CAT CTG GAG AAC ATC Ser Arg Ala Asn Thr Val Arg Ser Phe His His Glu Glu His Leu Glu Asn Ile
792 807 822 837 CCA GGG ACC AGT GAA AAC TCT GCT TTT CGT TTC CTC TTT AAC CTC AGC AGC ATC Pro Gly Thr Ser Glu Asn Ser Ala Phe Arg Phe Leu Phe Asn Leu Ser Ser Ile

6/8

Figure 2A

852 867 882 897 CCT GAG AAC GAG GTG ATC TCC TCT GCA GAG CTC CTC CGG GAG CAG GTG Pro Glu Asn Glu Val Ile Ser Ser Ala Glu Leu Arg Leu Phe Arg Glu Gln Val
912 927 942 GAC CAG GGC CCT GAT TGG GAA AGG GGC TTC CAC CGT ATA AAC ATT TAT GAG GTT Asp Gln Gly Pro Asp Trp Glu Arg Gly Phe His Arg Ile Asn Ile Tyr Glu Val
957 972 987 1002 ATG AAG CCC CCA GCA GAA GTG GTG CCT GGG CAC CTC ATC ACA CGA CTA CTG GAC MET Lys Pro Pro Ala Glu Val Val Pro Gly His Leu Ile Thr Arg Leu Leu Asp
1017 1032 1047 ACG AGA CTG GTC CAC CAC AAT GTG ACA CGG TGG GAA ACT TTT GAT GTG AGC CCT Thr Arg Leu Val His His Asn Val Thr Arg Trp Glu Thr Phe Asp Val Ser Pro
1062 1077 1092 1107 GCG GTC CTT CGC TGG ACC CGG GAG AAG CAG CCA AAC TAT GGG CTA GCC ATT GAG Ala Val Leu Arg Trp Thr Arg Glu Lys Gln Pro Asn Tyr Gly Leu Ala Ile Glu
1122 1137 1152 1167 GTG ACT CAC CTC CAT CAG ACT CGG ACC CAC CAG GGC CAG CAT GTC AGG ATT AGC Val Thr His Leu His Gln Thr Arg Thr His Gln Gly Gln His Val Arg Ile Ser
1182 1197 1212 CGA TCG TTA CCT CAA GGG AGT GGG AAT TGG GCC CAG CTC CGG CCC CTC CTG GTC Arg Ser Leu Pro Gln Gly Ser Gly Asn Trp Ala Gln Leu Arg Pro Leu Leu Val
1227 1242 1257 1272 ACC TTT GGC CAT GAT GGC CGG GGC CAT GCC TTG ACC CGA CGC CGG AGG GCC AAG Thr Phe Gly His Asp Gly Arg Gly His Ala Leu Thr Arg Arg Arg Ala Lys
1287 1302 1317
CGT AGC CCT AAG CAT CAC TCA CAG CGG GCC AGG AAG AAG AAT AAG AAC TGC CGS Arg Ser Pro Lys His His Ser Gln Arg Ala Arg Lys Lys Asn Lys Asn Cys Arg
1332(311) 1347 1362 1377 CGC CAC TCG CTC TAT GTG GAC TTC AGC GAT GTG GGC TGG AAT GAC TGG ATT GTG Arg His Ser Leu Tyr Val Asp Phe Ser Asp Val Gly Trp Asn Asp Trp Ile Val
1392 1407 1422 1437 GCC CCA CCA GGC TAC CAG GCC TTC TAC TGC CAT GGG GAC TGC CCC TTT CCA CTG Ala Pro Pro Gly Tyr Gln Ala Phe Tyr Cys His Gly Asp Cys Pro Phe Pro Leu
1452 1467 1482 GCT GAC CAC CTC AAC TCA ACC AAC CAT GCC ATT GTG CAG ACC CTG GTC AAT TCT Ala Asp His Leu Asn Ser Thr Asn His Ala Ile Val Gln Thr Leu Val Asn Ser
1497 1512 1527 1542 GTC AAT TCC AGT ATC CCC AAA GCC TGT TGT GTG CCC ACT GAA CTG AGT GCC ATC Val Asn Ser Ser Ile Pro Lys Ala Cys Cys Val Pro Thr Glu Leu Ser Ala Ile

1946 CTAGAGTCGA CGGAATTC 7/8

Figure 2B

1557 1572 TCC ATG CTG TAC CTG GAT GAG TAT GAT AAG GTG GTA CTG AAA AAT TAT CAG GAG Ser MET Leu Tyr Leu Asp Glu Tyr Asp Lys Val Val Leu Lys Asn Tyr Gln Glu 1617 (408) ATG GTA GTA GAG GGA TGT GGG TGC CGC TGAGATCAGG CAGTCCTTGA GGATAGACAG 1636 MET Val Val Glu Gly Cys Gly Cys Arg 1666 1676 1696 1726 1746 1756 1766 ACAGACTGCT TCCTTATAGC TGGACTTTTA TTTAAAAAAA AAAAAAAAA AATGGAAAAA ATCCCTAAAC 1776 1816 1826 1836 ATTCACCTTG ACCTTATTTA TGACTTTACG TGCAAATGTT TTGACCATAT TGATCATATA TTTTGACAAA 1866 1886 1896 1906 ATATATTAT AACTACGTAT TAAAAGAAAA AAATAAAATG AGTCATTATT TTAAAAAAAA AAAAAAAACT 1916



Figure 3

*	ACA Thr -40	AGA Arg	GAG Glu	TGC Cys	TCA Ser	AGA Arg -35	AGC Ser	TGT Cys	CCA Pro	AGG Arg	ACG Thr -30	GCT Ala	CCA Pro	CAG Gln	AGG Arg	48
CAG Gln -25	GTG Val	AGA Arg	GCA Ala	GTC Val	ACG Thr -20	AGG Arg	AGG Arg	ACA Thr	CGG	ATG Met -15	GCG Ala	CAC His	GTG Val	GCT Ala	GCG Ala -10	96
GGG Gly	TCG Ser	ACT Thr	TTA Leu	GCC Ala -5	AGG Arg	CGG Arg	AAA Lys	AGG Arg	AGC Ser 1	GCC Ala	GGG Gly	GCT Ala	GGC Gly 5	AGC Ser	CAC His	144
TGT Cys	CAA Gln	AAG Lys 10	ACC Thr	TCC Ser	CTG Leu	CGG	GTA Val 15	AAC Asn	TTC Phe	GAG Glu	GAC Asp	ATC Ile 20	GGC Gly	TGG Trp	GAC Asp	192
AGC Ser	TGG Trp 25	ATC Ile	ATT Ile	GCA Ala	CCC Pro	AAG Lys 30	GAG Glu	TAT Tyr	GAA Glu	GCC Ala	TAC Tyr 35	GAG Glu	TGT Cys	AAG Lys	GGC Gly	240
GGC Gly 40	TGC Cys	TTC Phe	TTC Phe	CCC Pro	TTG Leu 45	GCT Ala	GAC Asp	GAT Asp	GTG Val	ACG Thr 50	CCG Pro	ACG Thr	AAA Lys	CAC His	GCT Ala 55	288
ATC Ile	GTG Val	CAG Gln	ACC Thr	CTG Leu 60	GTG Val	CAT His	CTC Leu	AAG Lys	TTC Phe 65	ĊCC Pro	ACA Thr	AAG Lys	GTG Val	GGC Gly 70	AAG Lys	336
GCC Ala	TGC Cys	TGT Cys	GTG Val 75	CCC Pro	ACC Thr	AAA Lys	CTG Leu	AGC Ser 80	CCC Pro	ATC Ile	TCC Ser	GTC Val	CTC Leu 85	TAC Tyr	AAG Lys	384
GAT Asp	GAC Asp	ATG Met 90	GGG Gly	GTG Val	CCC Pro	ACC Thr	CTC Leu 95	AAG Lys	TAC Tyr	CAT His	TAC Tyr	GAG Glu 100	GGC Gly	ATG Met	AGC Ser	432
GTG Val	GCA Ala 105	GAG Glu	TGT Cys	GGG Gly	TGC Cys	AGG Arg 110	TAG	PATCI	rgc (CTGC	GG					. 470

I. CLASSIFI	CATION OF SUBJE	CT MATTER (if several classification syn	nbols apply, indicate all) ⁶	
		Classification (IPC) or to both National Cla	ssification and IPC	
Int.Cl.	5 C12N15/1	2; C12P21/02;	A61K37/02	
II. FIELDS	SEARCHED			
		Minimum Documen	nation Searched?	
Classification	on System		Jassification Symbols	
Int.Cl.	5	CO7K ; C12N ;	A61K	
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		9		
	MENTS CONSIDER	ED TO BE RELEVANT ⁹ ocument, ¹¹ with indication, where appropria	to of the relative pressures 12	Relevant to Claim No.13
Category °	Citation of D	ocument, " with indication, where appropria	te, or the relevant passages	
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A	SCIENCE vol. 87 US pages 9 CELESTE transfo members purifie	INGS OF THE NATIONAL ACS OF USA, no. 24, December 1990 843 - 9847 A.J. ET AL. 'Identification growth factor bet present in bone-inducted from bovine bone's whole document	, WASHINGTON cation of a family	1-18
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, A	WO,A,9 118 098 (GENETICS INSTITUTE, INC.) 28 November 1991 cited in the application see the whole document	1-18
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INTERNATIONAL SEARCH REPORT

PCT/US 92/05374

Box I	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)	
This int	ernational search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:	
1. X	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely: Remark: Although claims 16, 18 are directed to a method of treatment of the human/animal body the search has been carried out and based on the alleged affects of the compound/composition.	
2.	Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:	
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).	
Box I	I Observations where unity of invention is lacking (Continuation of item 2 of first sheet)	
This I	nternational Searching Authority found multiple inventions in this international application, as follows:	
1		
1. [As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.	
2.	As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.	
3. [As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:	•
4. [No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:	
Ren	mark on Protest The additional search fees were accompanied by the applicant's protest.	
	No protest accompanied the payment of additional search fees.	



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Patent document cited in search report	Publication date	F	atent family member(s)	Publication date	
₩0-A-9011366	04-10-90	US-A- AU-A- CA-A- EP-A- JP-T-	5106748 5357790 2030518 0429570 3505098	21-04-92 22-10-90 29-09-90 05-06-91 07-11-91	
WO-A-9118098	28-11-91	None		p-23-43-47-48-4-	

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